



# Greater Amberjack Abundance, Distribution, and Movement in U.S. Waters in the South Atlantic and Gulf of Mexico

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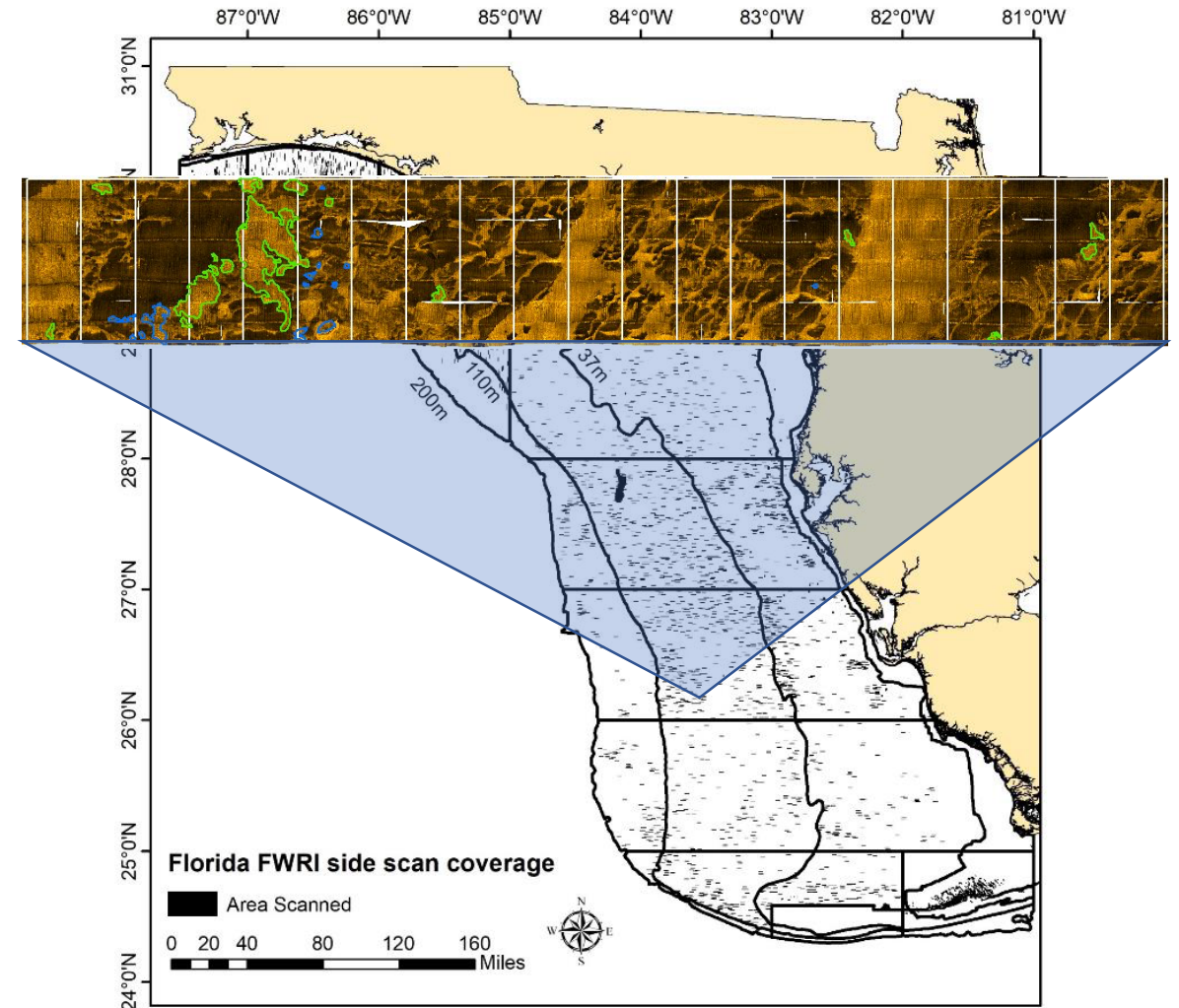


# Specific objectives

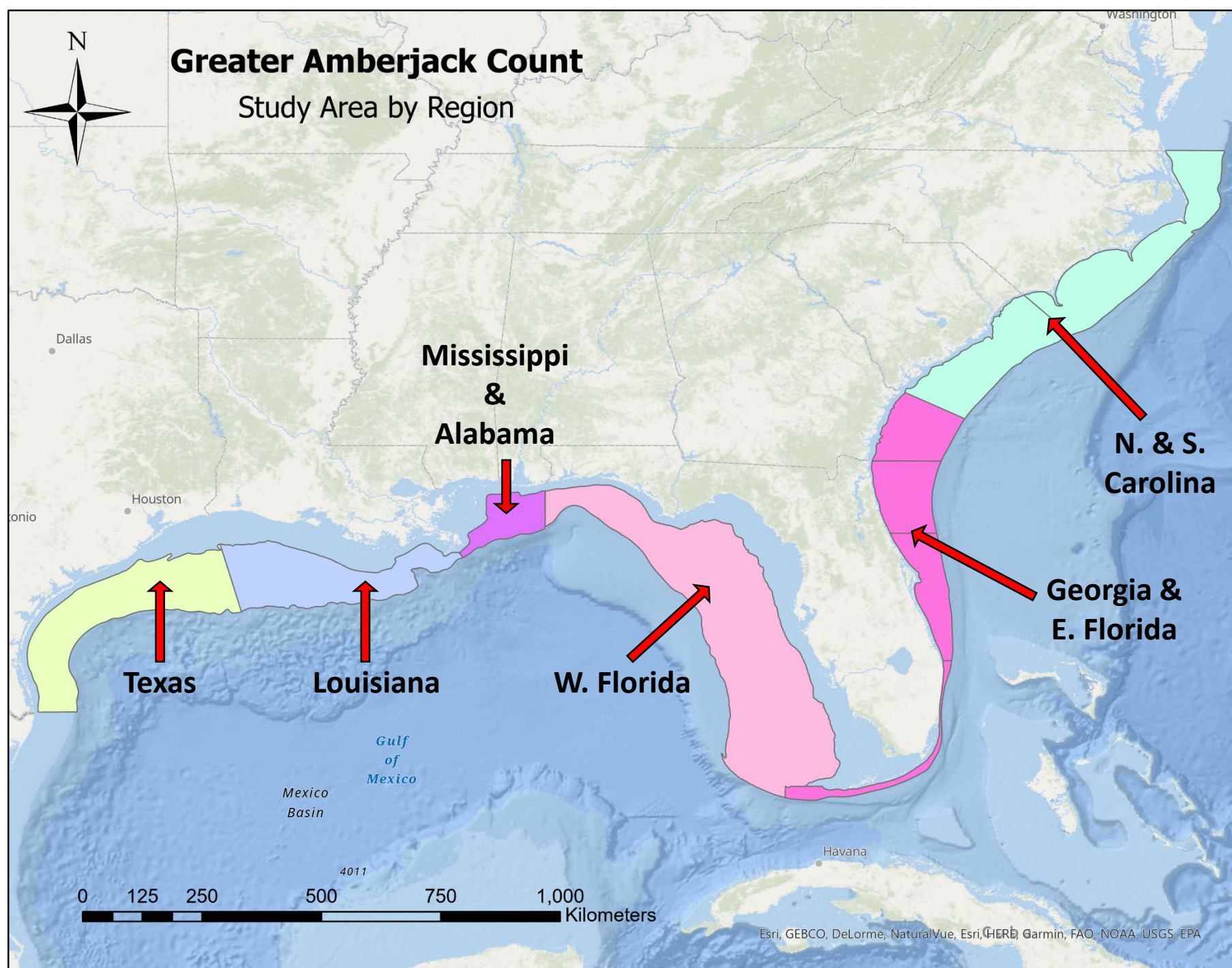
1. Synthesize existing bottom habitat observations
2. Synthesize existing abundance data - catch data and stakeholder knowledge
3. Comprehensive study to estimate regional, habitat-specific absolute abundance using video and hydroacoustics
4. Determine movement and connectivity using acoustic telemetry, conventional tagging, and genetic markers
5. Assess efficacy of eDNA to determine presence/relative abundance of GAJ and related species
6. Update biological information across study region
7. Engage in outreach to facilitate stakeholder input and communicate results

# Objective 1: Synthesize habitat data

- No existing comprehensive maps for entire region
- Existing sources of habitat data
  - partial coverage
  - variable resolution
- Compile existing habitat data into comprehensive GIS product across GoM-SA region
- Inform sampling design, and ultimately, final estimates



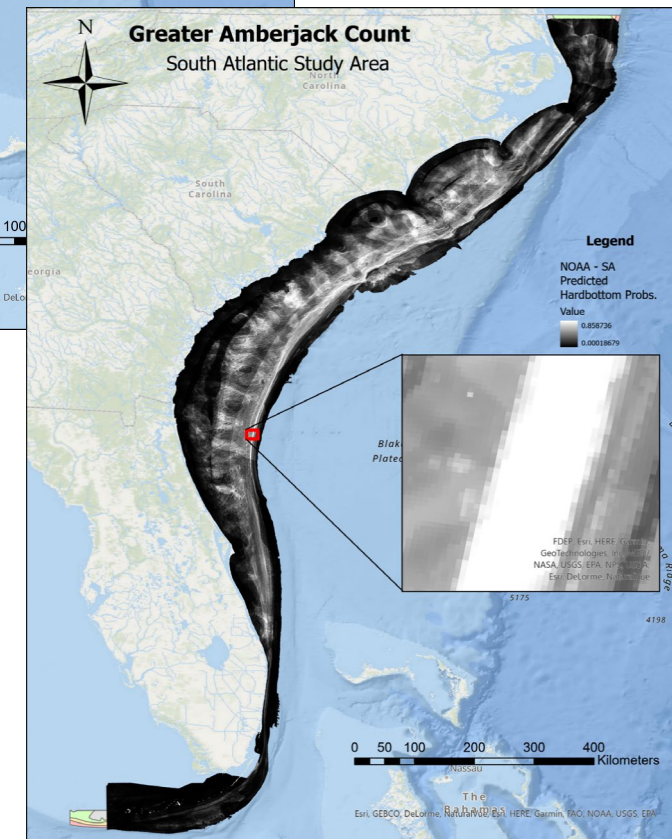
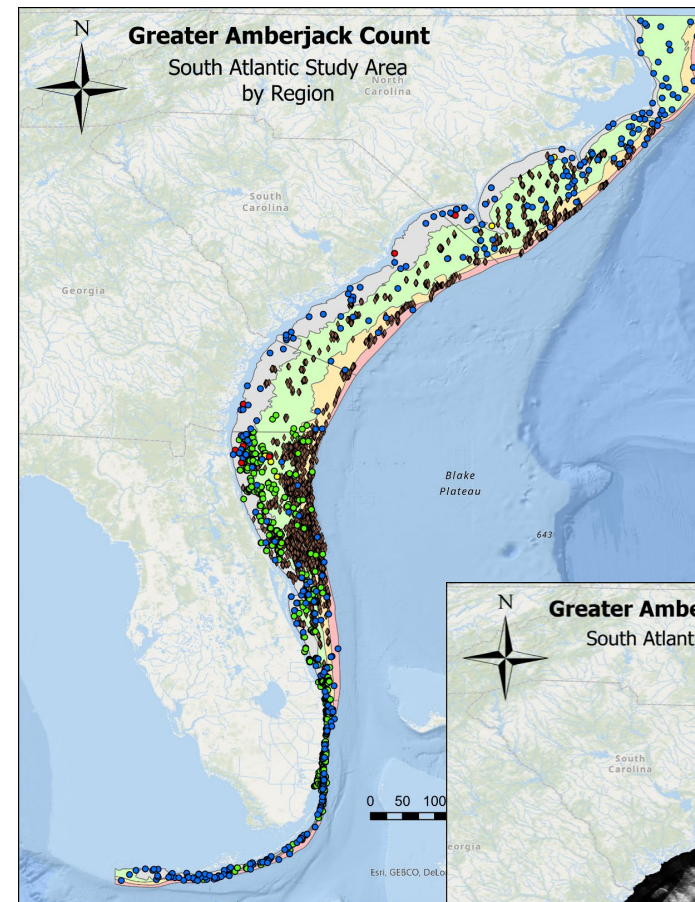






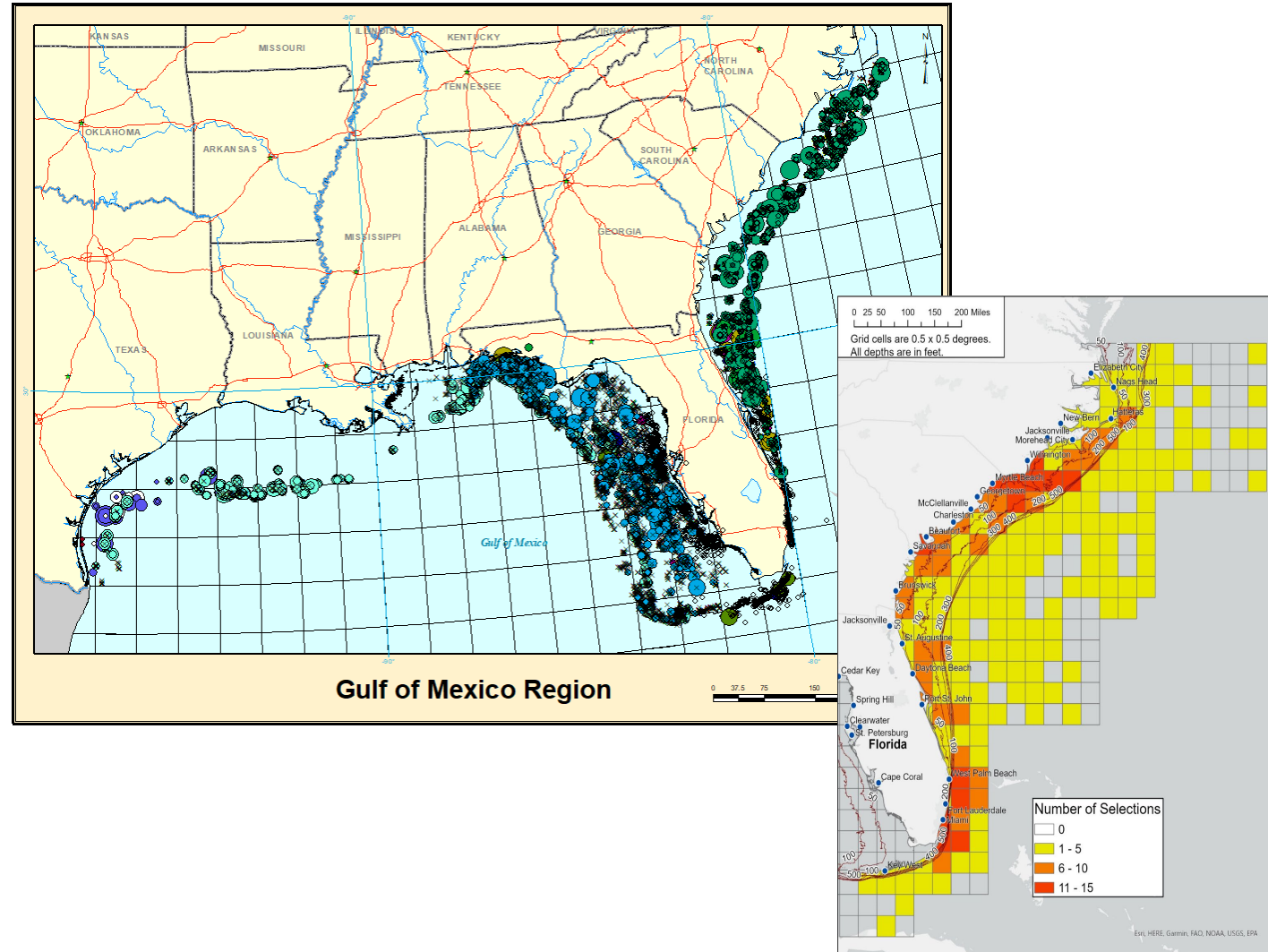
# Habitat synthesis: South Atlantic

- List of artificial reef locations/types/sizes (NOAA-ENC, FWRI)
- List of known natural reef point-locations (NOAA-SERFS, FWRI)
- Location and extent info for natural-reefs comes from prob. models (NCCOS)
- No scaleable habitat map products



# Objective 2: Synthesize abundance data

- Existing fishery dependent and fishery independent catch data
  - SERFS, G-FISHER, Project PIs, Observer programs (FL)
- Existing stakeholder knowledge (LEK)
- Inform expectations in terms of presence/absence, relative abundance and variance
  - Priors for Bayesian abundance models
  - More efficient sample design



# Objective 3: Estimate absolute abundance

- Abundance sampling methods
- Sample design and framework
- Calibration of gears

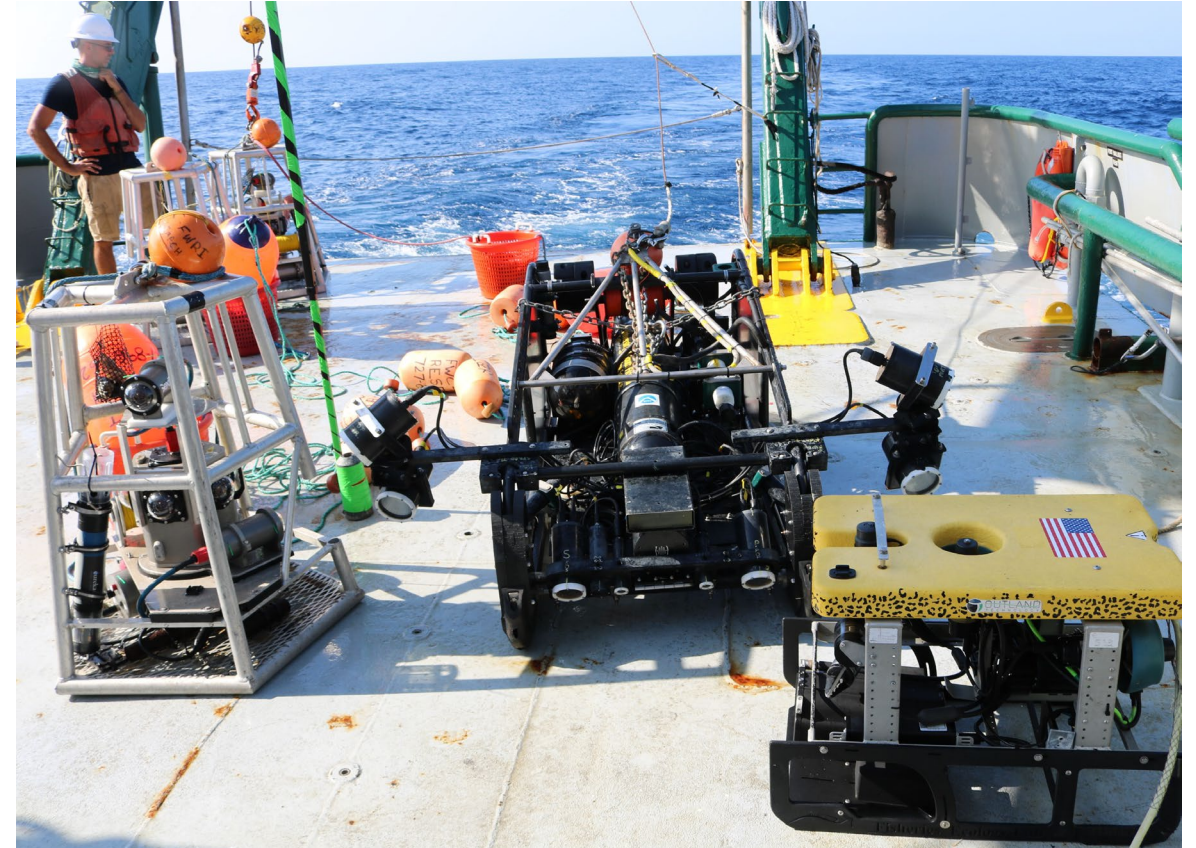


# Abundance sampling methods

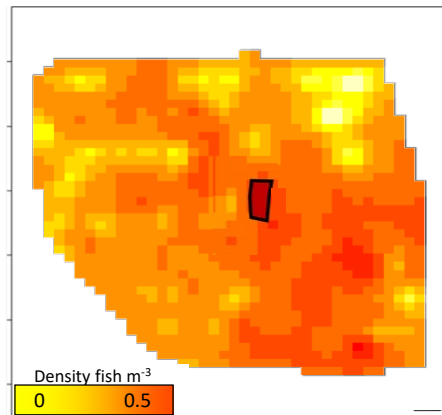
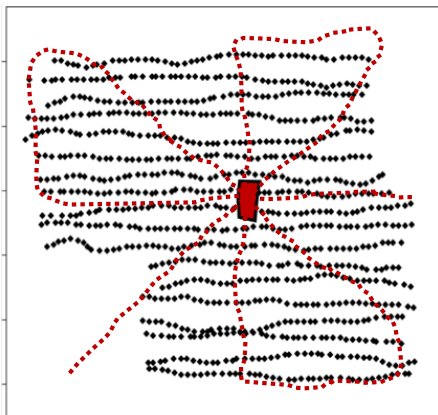
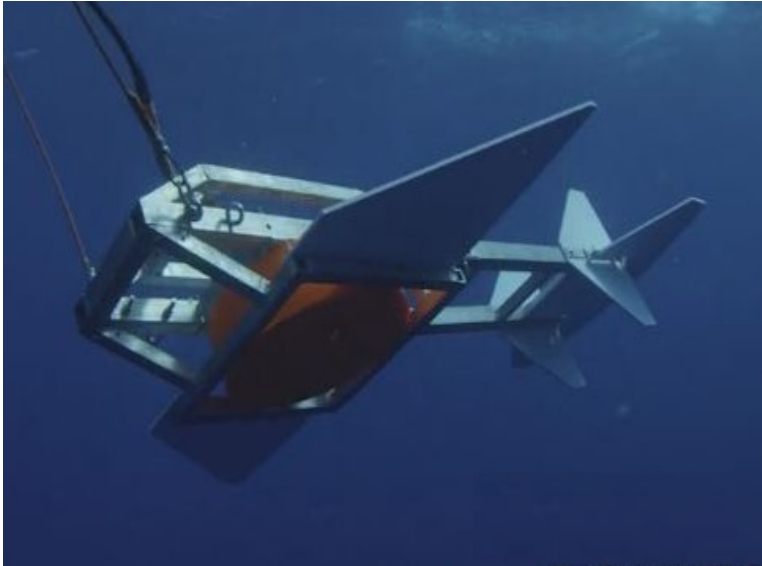
- Core approach: combine video (stationary, ROV, and towed) and active acoustics to measure density of GAJ
- Specific type of video is habitat- and region-specific due to advantages of each gear type
  - E.g. towed cameras effective for sampling large swathes of low-relief habitat, ROV effective for sampling high-relief artificial habitat
- Assess efficacy of emerging eDNA technologies
- Gears calibrated to each other and to a “ground-truth” abundance metric (Lincoln-Peterson estimate from VPS array)

# Video

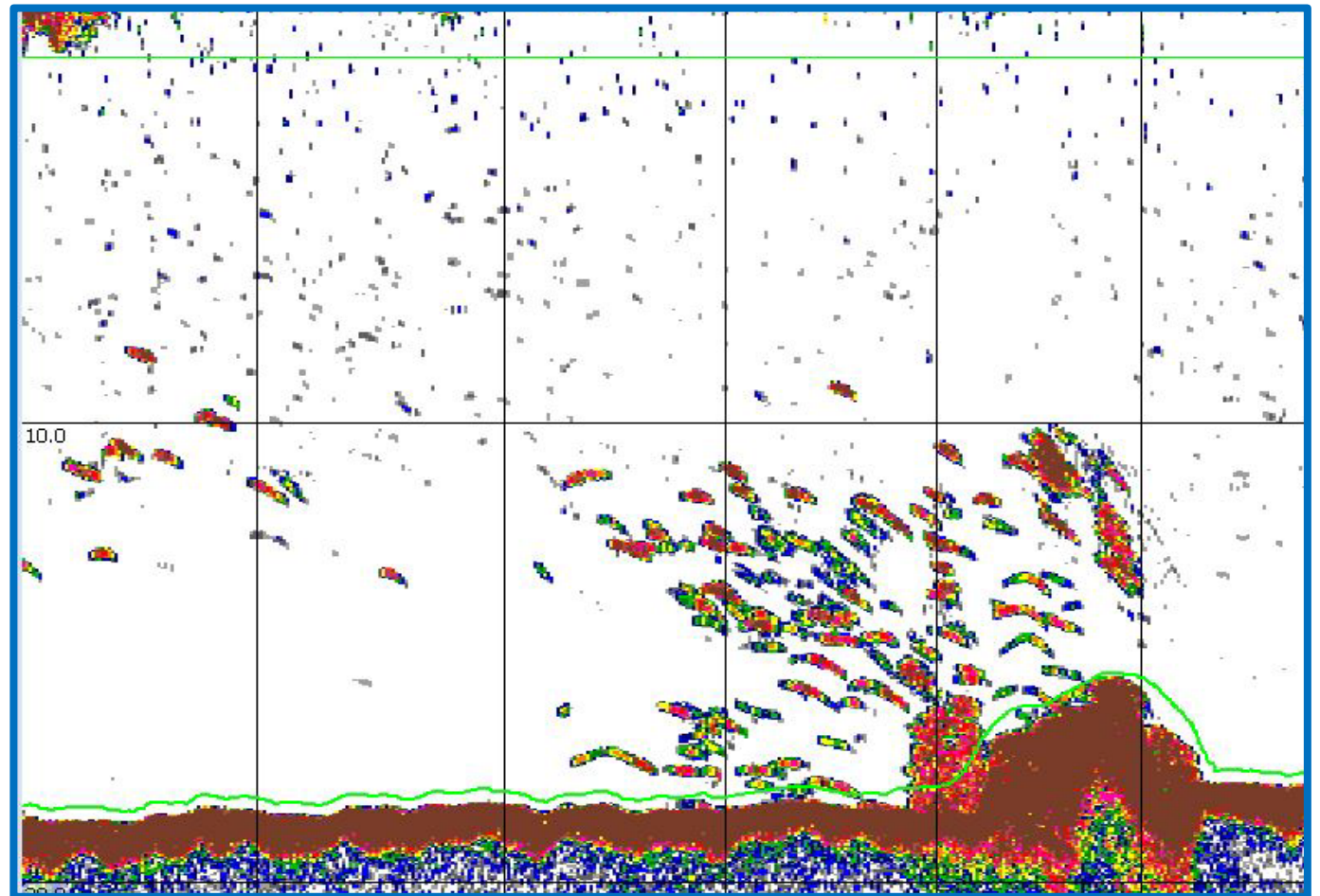
- Video types for different habitats
  - Baited drop cameras – artificial and natural reefs, all regions
  - ROV mounted cameras – artificial and natural reefs, GoM regions
  - Towed cameras – uncharacterized bottom, all regions
- Dedicated efforts to understand potential biases and how they influence probability of detection:
  - Attraction/avoidance
  - Influence of bait
  - Enumeration methods
  - Identification difficulties
- Calibration studies and coupling with active acoustics help to address these



# Active acoustics



Depth



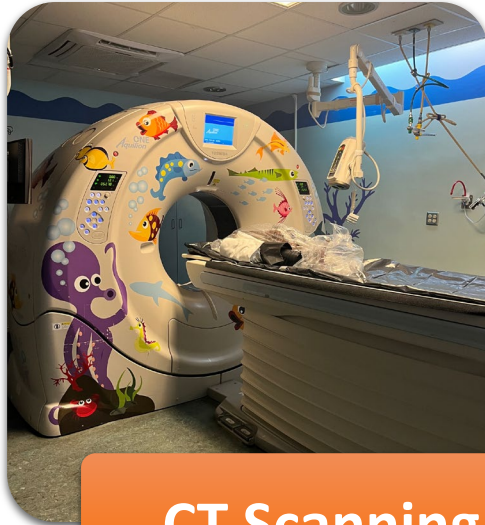
Distance along transect



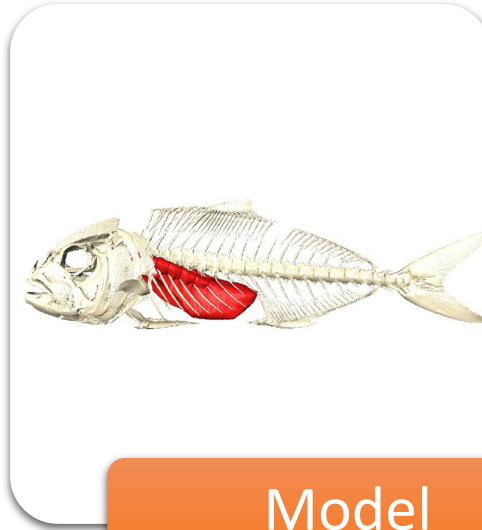
# Active acoustics: identifying Amberjack



**GAJ Collection**



**CT Scanning**



**Model  
Processing**



**Backscatter  
Modeling**

# Example Output

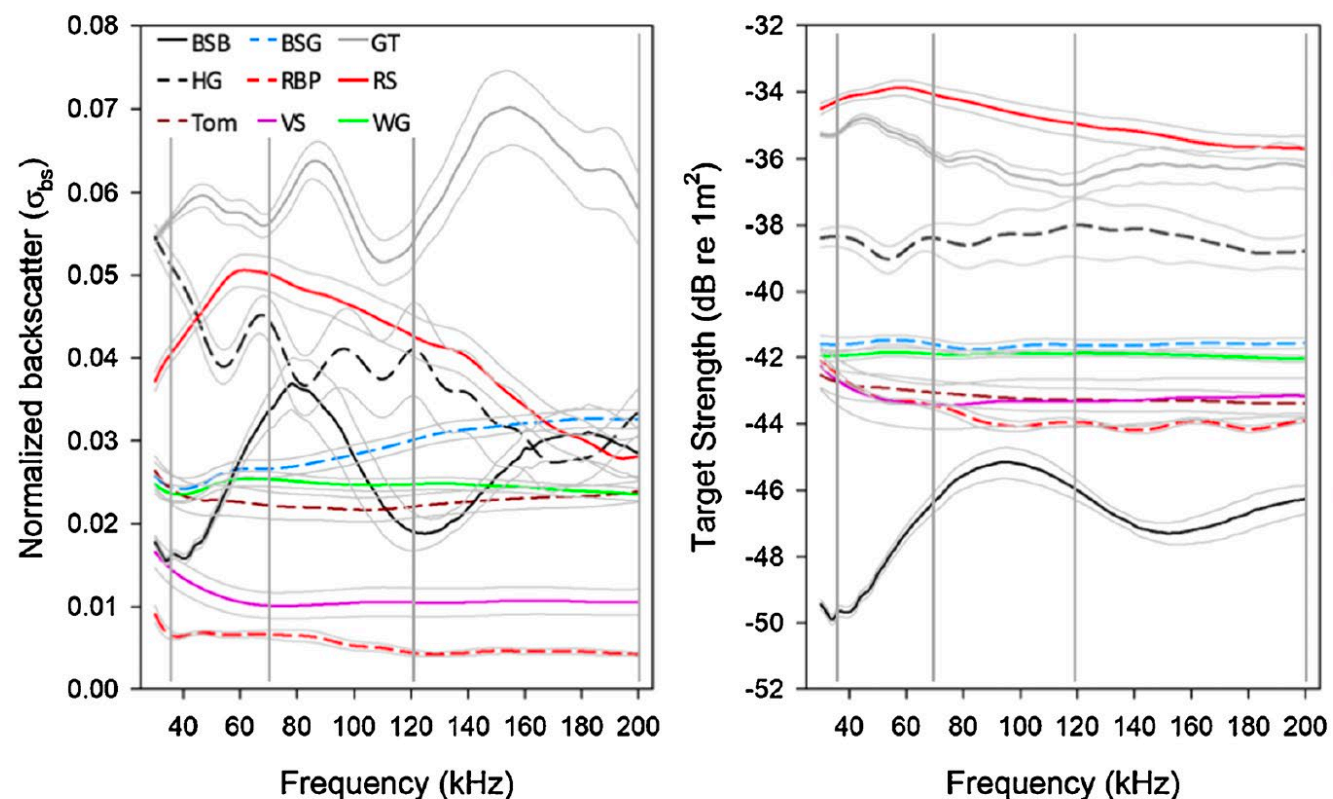
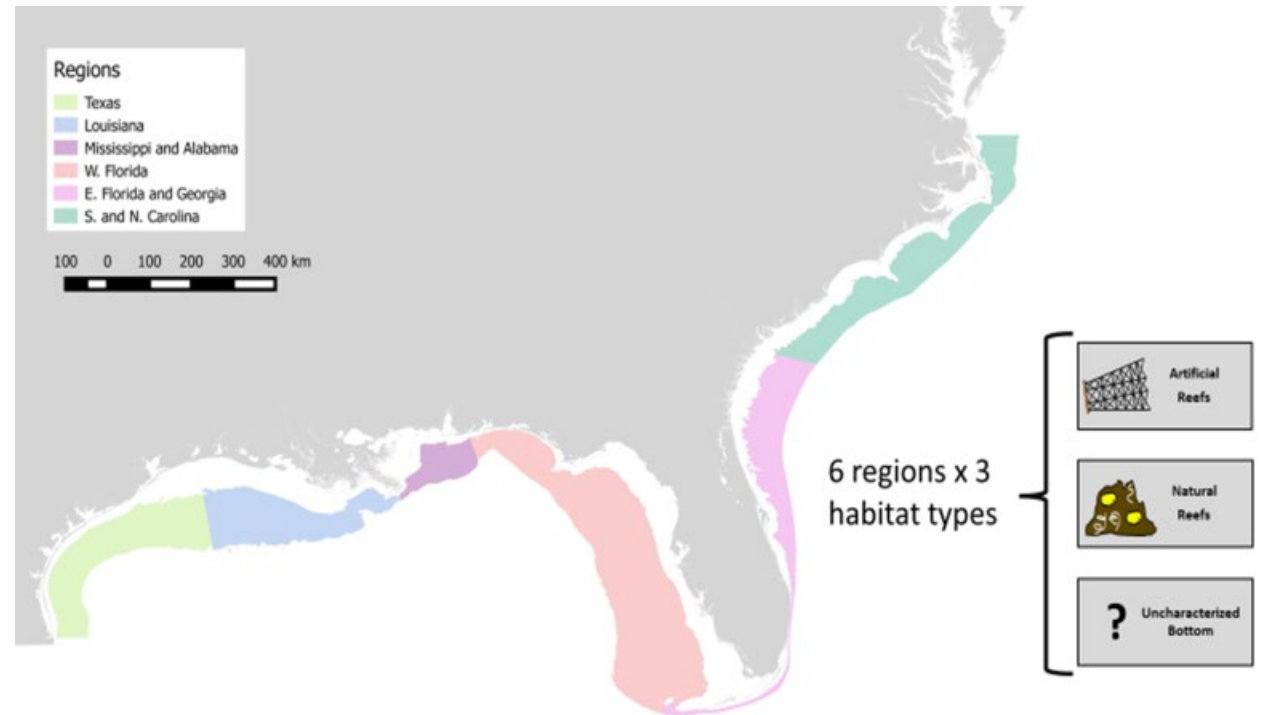


Fig. 6. Species specific averaged  $\sigma_{bs}$  response (m<sup>2</sup>) with 95 % confidence intervals (broken lines) around the mean (solid line). Vertical reference lines represent nominal operating frequencies in fisheries acoustics (38, 70, 120, and 200 kHz).

# Sample design and framework

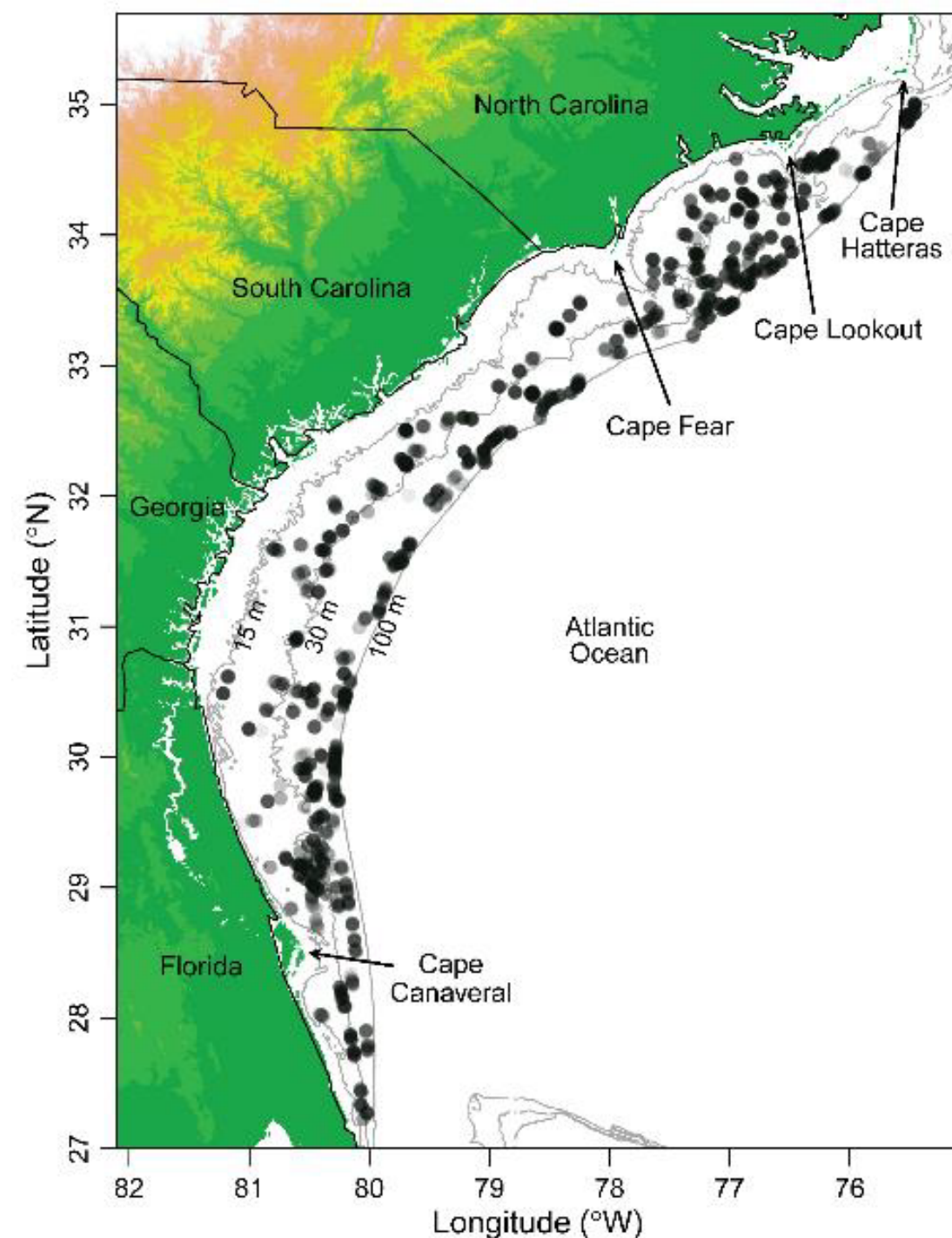
- Initial default (minimum) sample design is based on stratified random or cluster sampling by...
  - Region (TX, LA, MS-AL, West FL, East FL-GA, SC-NC)
  - Habitat type (artificial structure, natural structure, uncharacterized bottom)





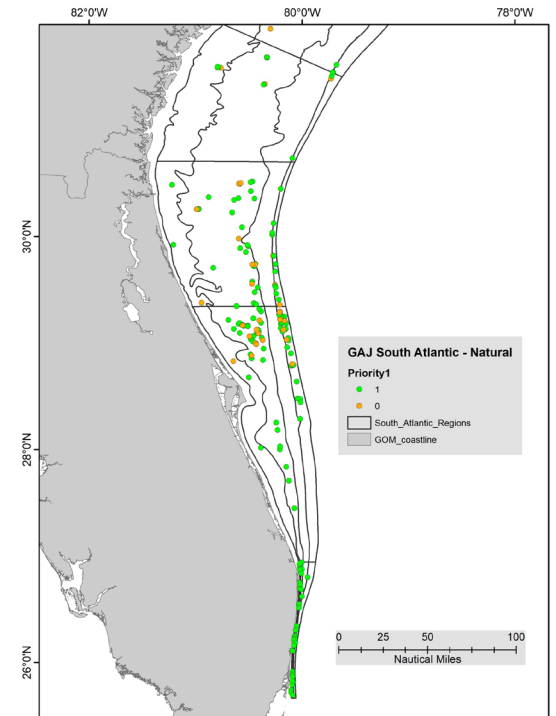
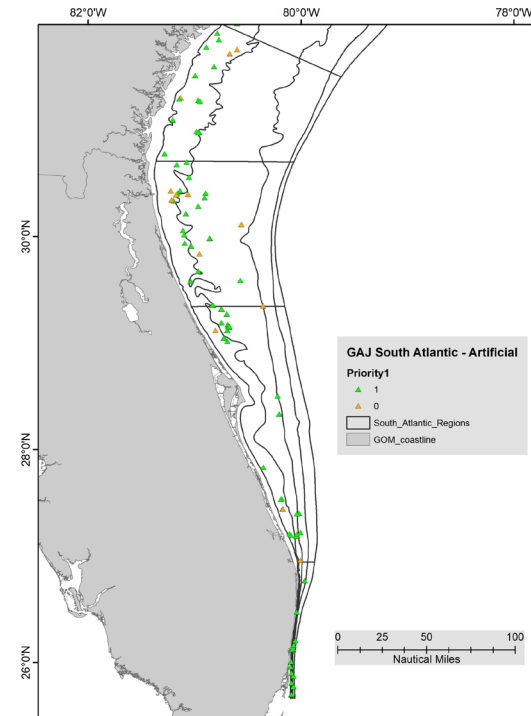
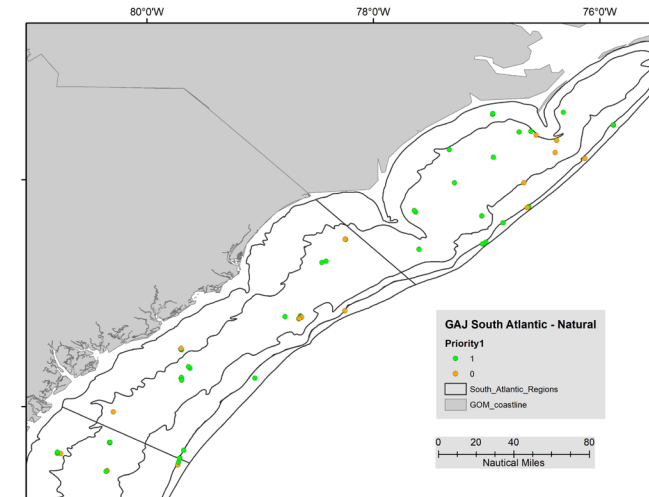
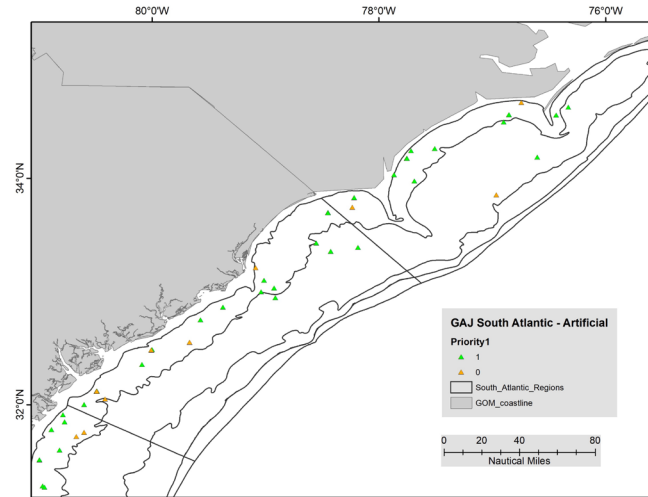
# Sample design: South Atlantic artificial and natural reefs

- Leverage SERFS (trap mounted cameras)
  - Known natural reef point-locations
  - Simple random sample from list of known natural reef point-locations
  - Does not cover artificial habitat
  - Cameras are depth limited
  - Does not cover SE FL



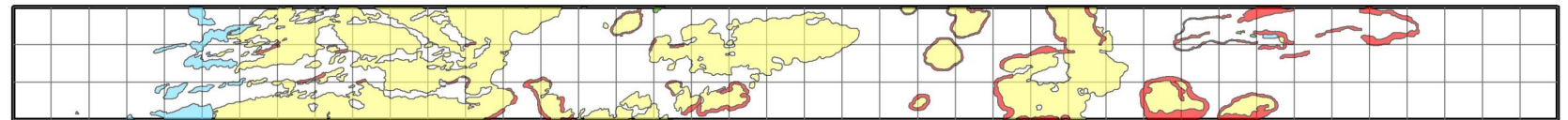
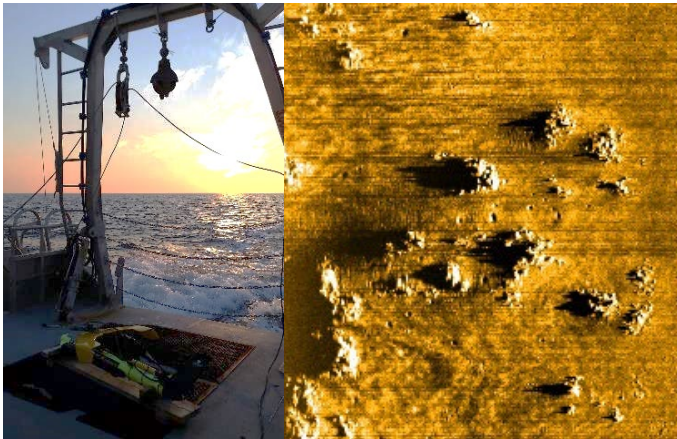
# Sample design: South Atlantic artificial and natural reefs

- S-BRUV + echosounder
  - Known natural and artificial reefs
  - Random sampling of point-locations stratified by region [five levels] and depth [three levels]
  - Will cover all depths, but extra effort in deeper waters and in SE FL where SERFS coverage is lacking



# South Atlantic – FWRI Habitat Mapping Plan

- Standardized mapping surveys at subset of natural reef sampling sites:
  - Klein 3900 SSS at 445 kHz
  - Survey orientation contingent on current:
    - Typically perpendicular to coast – may have to adjust in high N-S current areas
  - Aim to map ~ 30 – 40% of sampling sites (N ~ 60 surveys)
  - Centered on selected sampling point (often cover multiple potential sampling points)
  - Provide estimates of reef area and/or number of features



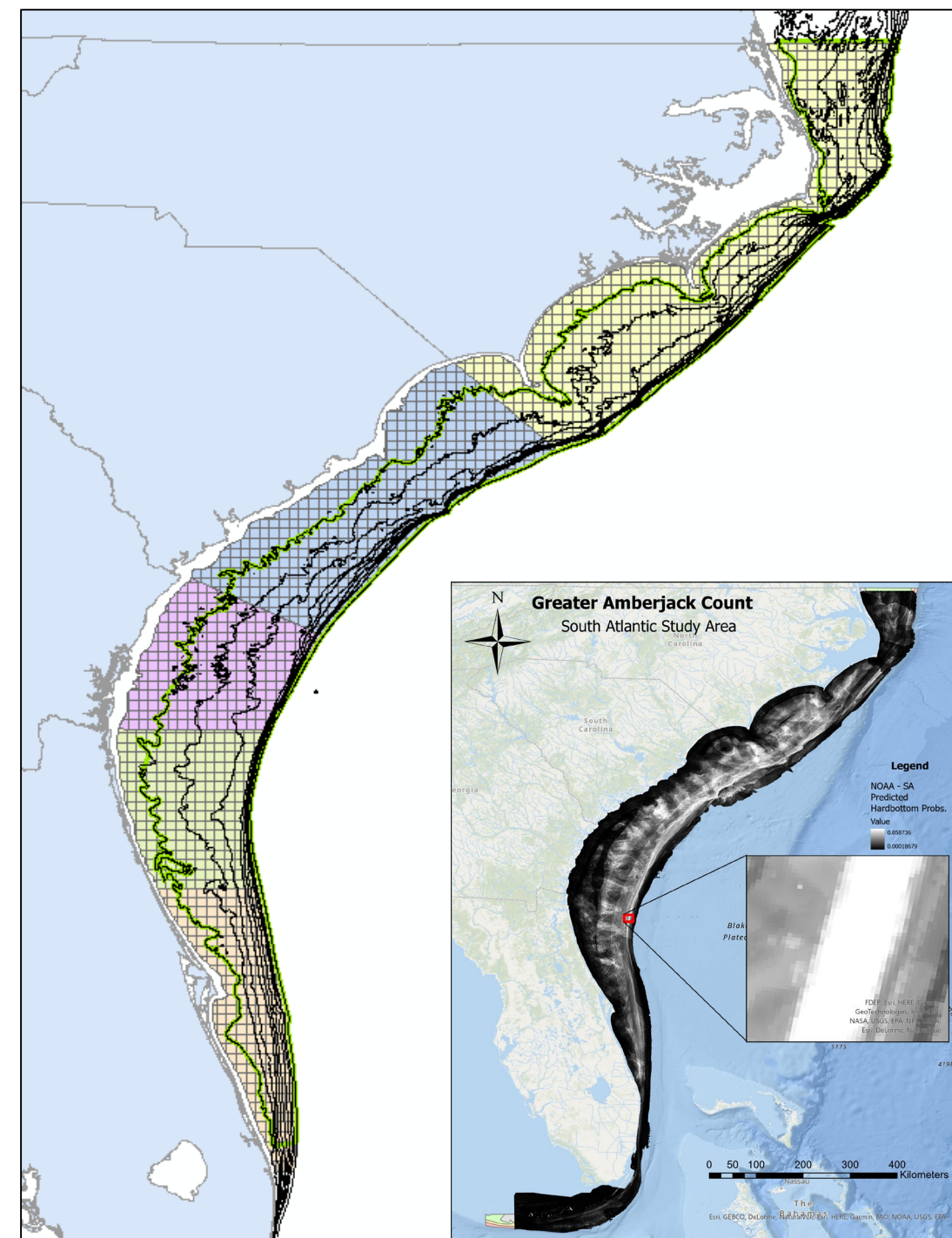
## Geoforms

- Geologic | Ledge
- Geologic | FragmentedHB
- Geologic | Pinnacle
- Geologic | FlatHB



# Sample design: South Atlantic uncharacterized

- C-BASS + echosounder
- Random sampling stratified by region [two levels] and depth [two levels]
- Multibeam mapping provides estimate of unknown natural and artificial reefs (potentially validation of NCCOS model)



# Calibration of gears and methods

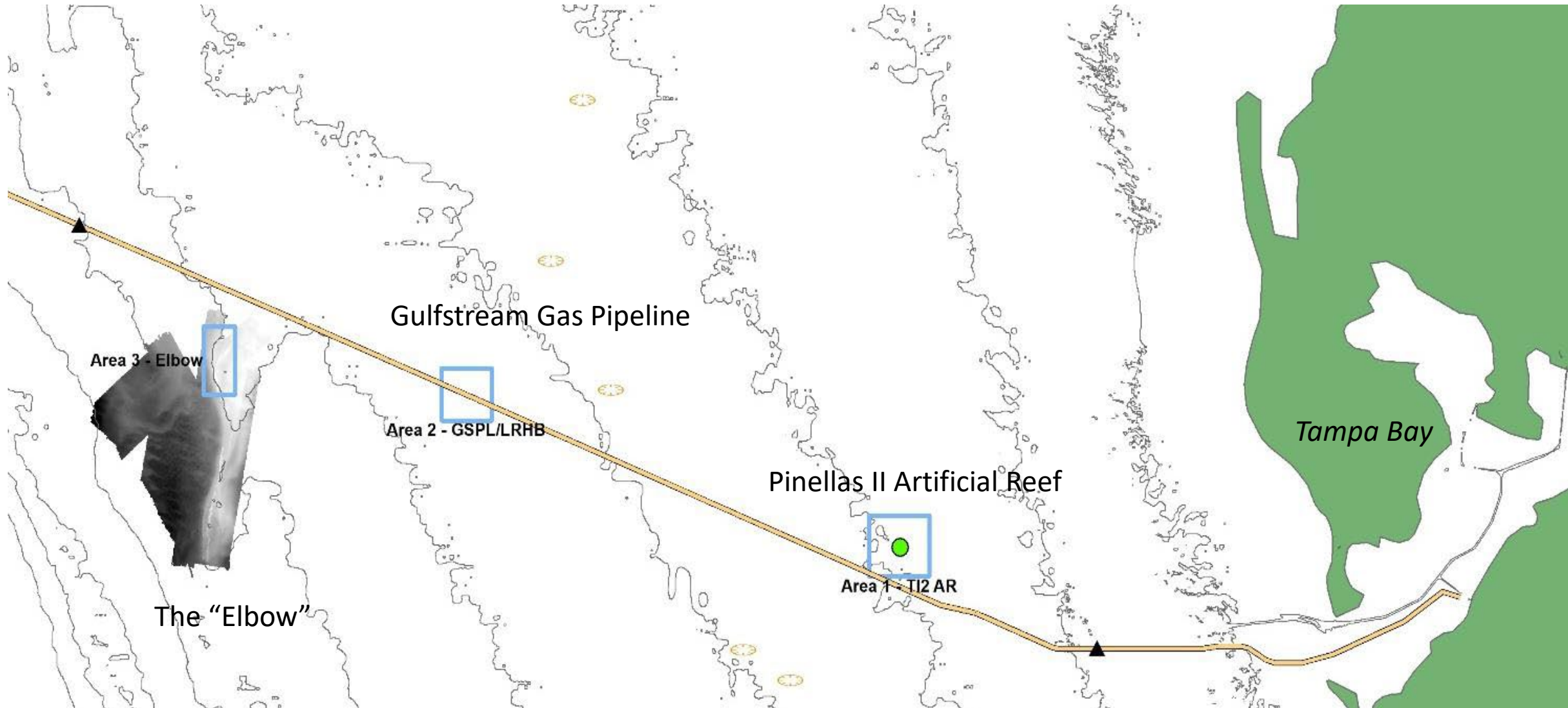
- Comparisons of camera gears
  - Baited vs. un-baited stationary cameras
  - Stationary vs. ROV
  - Stationary vs. towed
  - ROV vs. towed
- Active acoustics vs. all camera gears
- All gears (cameras and active acoustics) vs. ground-truth (Lincoln-Peterson estimate of abundance within a VPS array)
- eDNA vs. all other gears

# Calibration: Florida (May 4-10, 2022)

## Objectives:

- Test gears
- Deploy multiple gears same-time, same-place
- Compare results among gears
- Estimate calibration factors

# Survey Areas

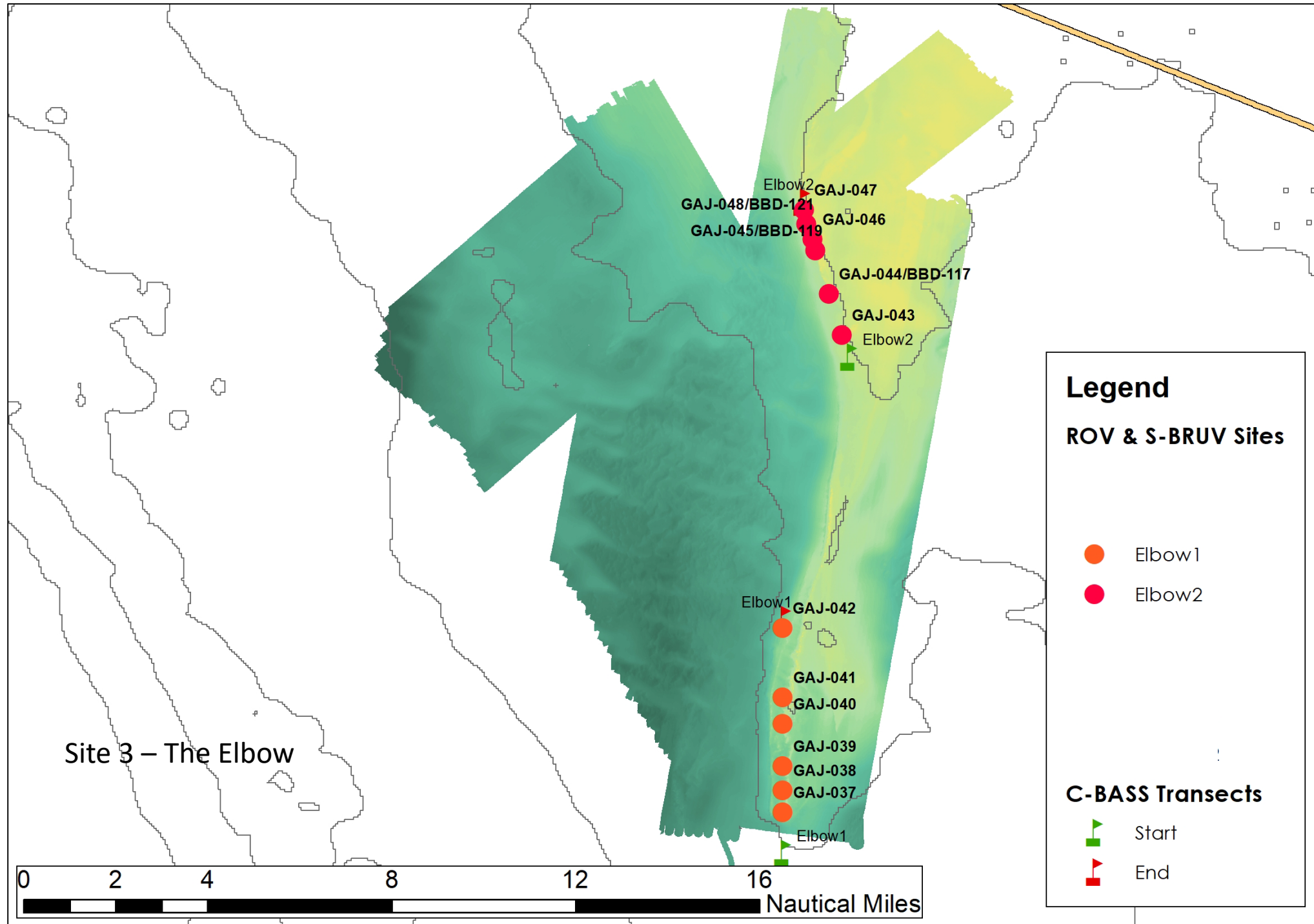




# Sampling protocol

- Each gear sampled every day, order randomized
- Echosounder running continuously
- C-BASS not deployed on artificial reef site

Site	1st Gear	2nd Gear	3rd Gear
Site 1 (Artificial)	S-BRUV	<del>C-BASS</del>	ROV
Site 2 (Artificial)	<del>C-BASS</del>	ROV	S-BRUV
Site 3 (Pipeline)	ROV	C-BASS	S-BRUV
Site 4 (Pipeline)	ROV	S-BRUV	C-BASS
Site 5 (Elbow)	S-BRUV	C-BASS	ROV
Site 6 (Elbow)	ROV	C-BASS	S-BRUV

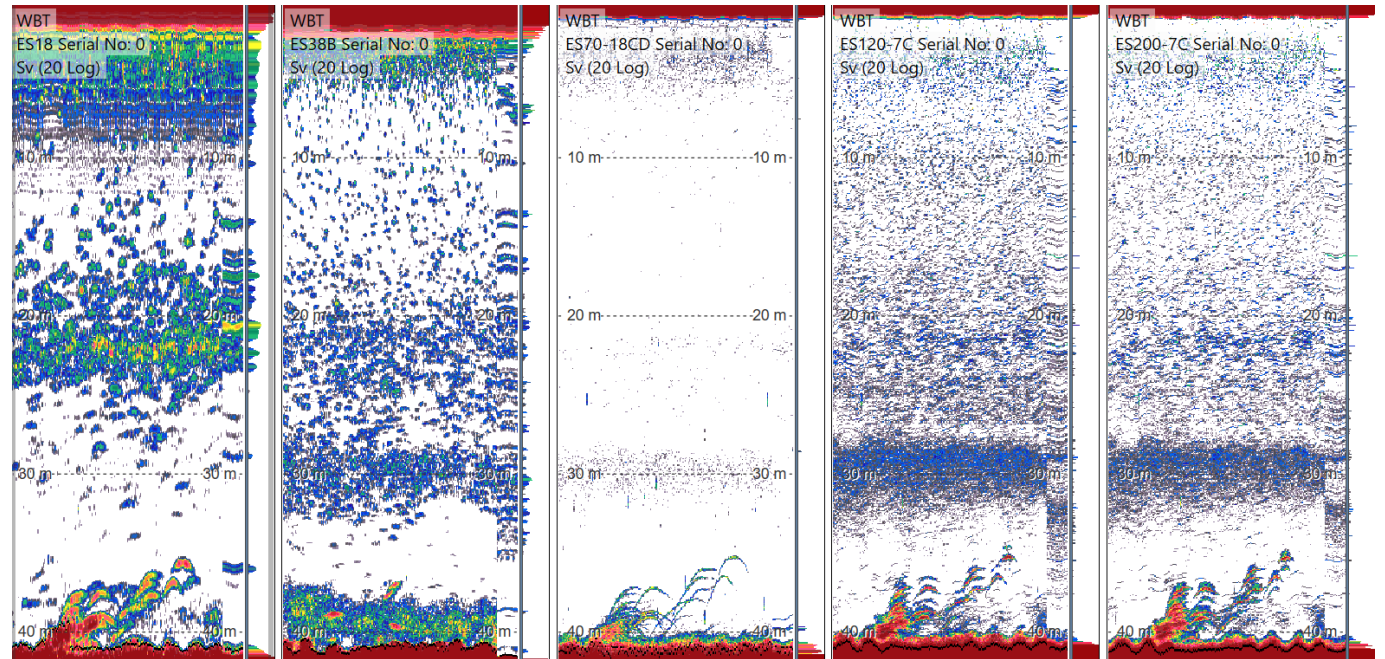


# Preliminary results

- *Seriola* species (Greater amberjack, Almaco Jack, Banded rudderfish) seen at all locations
- All gear systems functioned as designed/expected
- Water visibility generally good to excellent
- ROV and C-BASS (except habitat, 75% complete) video reads are done, S-BRUVs are in progress; EK analyses done

# Preliminary results

- ROV
  - *S. dumerili*: 99; *Seriola* spp.: 3
  - Many mixed schools of *Seriola*
  - Highest counts on artificial reefs, much lower on pipeline and Elbow (flat hardbottom, small ledges)
- C-BASS
  - *S. dumerili*: 4; *Seriola* spp.: 7
  - Linking fish to habitat observations
- Echosounder
  - many fish observed, but not categorized to species level
  - working out *Seriola* acoustic signatures
  - Application of abundance models to “always-on” track data problematic
- S-BRUV
  - Video reads not finished





# Next steps

- Finish S-BRUV video reads
- Compare S-BRUV to ROV counts
- Parse C-BASS data for overlap with other camera gears and compare
- Test alternative echosounder survey patterns at next calibration

# Main takeaways

- Water clarity was good and once video reads are completed, we expect to have data to inform calibration factor estimates among camera gears
- Always-on echosounder of limited value for calculating areal abundance; need to use patterned (parallel lines or flower) survey for spatial models of abundance

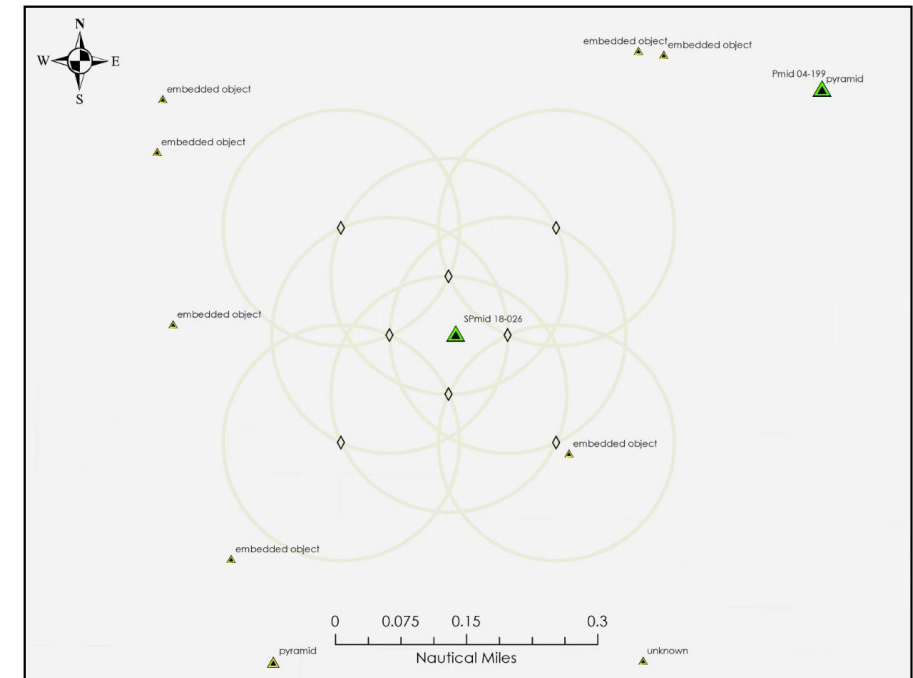
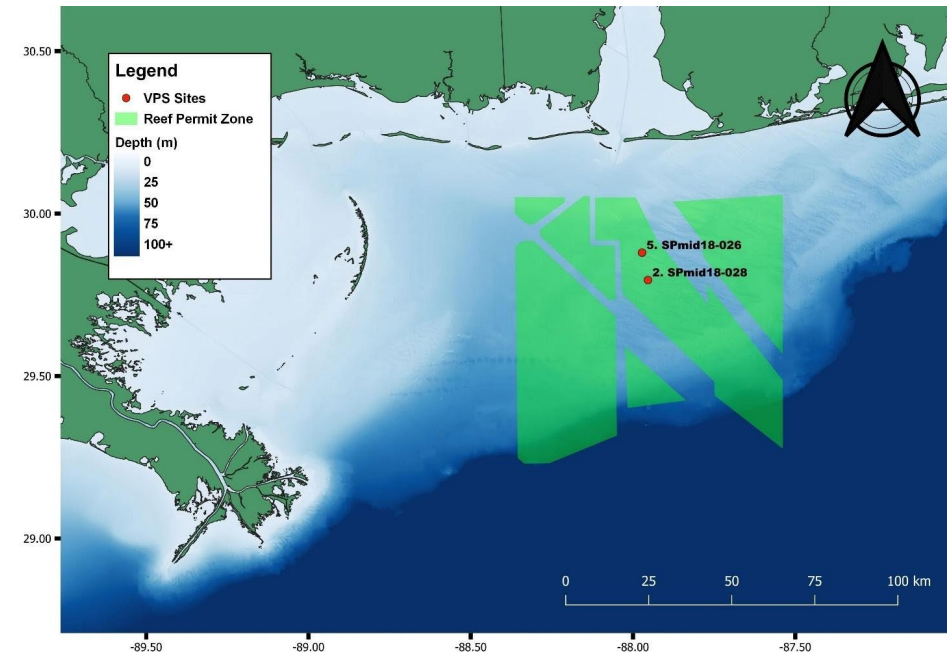
# Calibration: Mississippi/Alabama (Aug. 21-Sept. 2)

## Objectives

- Establish two VPS arrays with acoustically tagged *S. dumerili*
- Deploy multiple camera gears near concurrently in arrays
- Deploy active acoustics using different survey patterns (parallel lines, flower) and frequencies near concurrently in arrays
- Use VPS triangulated positions in combination with observations of tagged and untagged *S. dumerili* from camera gears to calculate Lincoln-Peterson abundance estimates as “ground truth”
- Use VPS triangulated positions to quantify behavioral changes in response to gear deployments
- Trial eDNA sample collection and assay efficacy at sites with known *S. dumerili*

# Methods

- VPS arrays deployed at two sites
- “Super pyramids” 25’ tall, 15’ base
- 8 receivers per site
- Min range ~ 250m
- Min coverage area ~20 hectares
- Acoustic + dart tags: 18 & 20 fish
- Dart tags: 5 & 3 fish





# Methods

- Two vessels
  - Escape:
    - ROV (AL/MS)
    - Drop Cam (Western GoM)
    - Active acoustics (All regions)
  - Wilson:
    - Trap Cam (SERFS)
    - S-BRUV (SA and FL)
    - eDNA (AL/MS)



# Methods

- One site designated as primary each day (alternate days)
- All gears deployed at primary site, with opportunistic deployments of “Wilson” gears at secondary site
- Vessel, gear order randomized each day except eDNA (before, after, and between other gears)

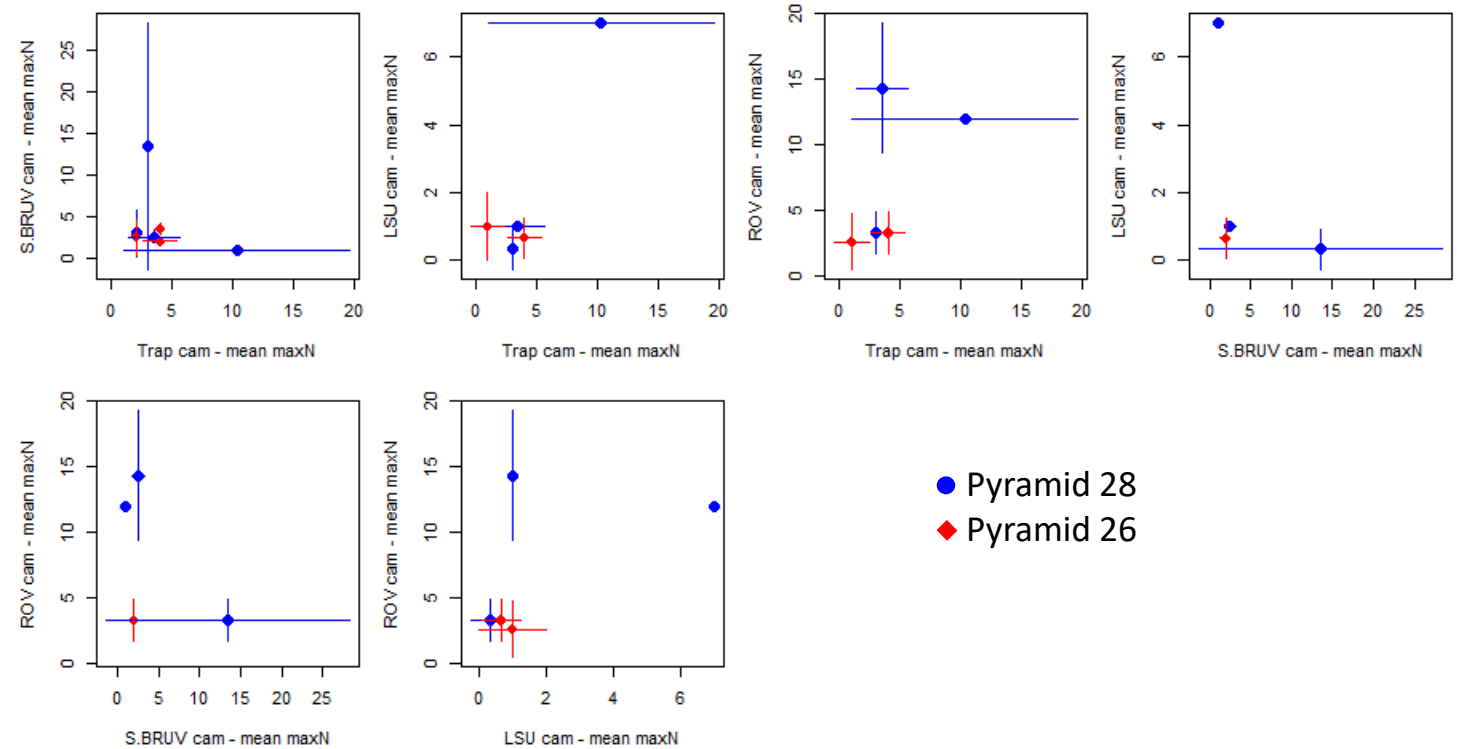
Sampling Allocation: 2-2.5 hr transit, 2 hrs/gear, 1.5 hr H-calib				
Vessel: Time	Wilson	Escape	Wilson	Escape
5:00				
5:30	Transit			
6:00				Transit
6:30				
7:00	DNA			
7:30				
8:00	G1			
8:30			Transit	G3
9:00				
9:30	DNA	Transit		
10:00				
10:30	G2			
11:00			2nd Site	G4
11:30		H-Calib		
12:00	DNA			G5
12:30			DNA	
13:00		G3	G1	
13:30	2nd Site			H-Calib
14:00				
14:30		G4	DNA	
15:00				
15:30	Transit		G2	Transit
16:00				
16:30		G5		
17:00			DNA	
17:30				
18:00			Transit	
18:30				
19:00		Transit		
19:30				
20:00				
20:30				
21:00				

# Preliminary results: camera gears

- Bait
  - Half of the S-BRUV drops were baited with the other half unbaited
  - No obvious difference in counts
- Proximity to reef
  - Half of the S-BRUV drops and half of the Trap Cam drops were near the reef (within 20 m) and half were far from the reef (~100 m away)
  - Near counts were substantially higher than far counts (mostly zeros)
- Time period
  - For some gears (Drop Cam, Trap Cam, S-BRUV), separate maxN counts were made for different periods over the deployment
  - Descent period had higher but more variable counts than bottom and ascent periods
  - Ascent period had lowest counts (mostly zeros)

# Preliminary results: camera gears

- Location
  - All gears had higher counts on Pyramid 28 than Pyramid 26
- Camera gear comparisons
  - ROV counts were generally higher than counts from other camera gears
  - Other than a general trend of higher counts on Pyramid 28, there were no strong correlations among camera gears
  - We believe that this will resolve with more concurrent samples at a larger number of sites

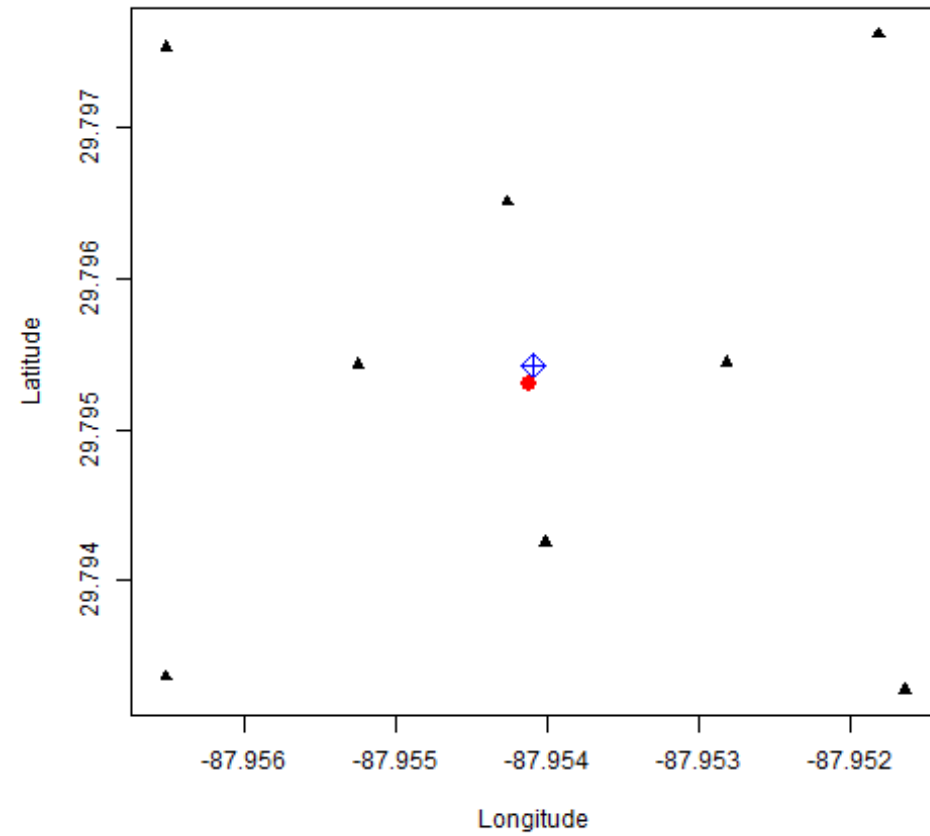




# Preliminary results: VPS array

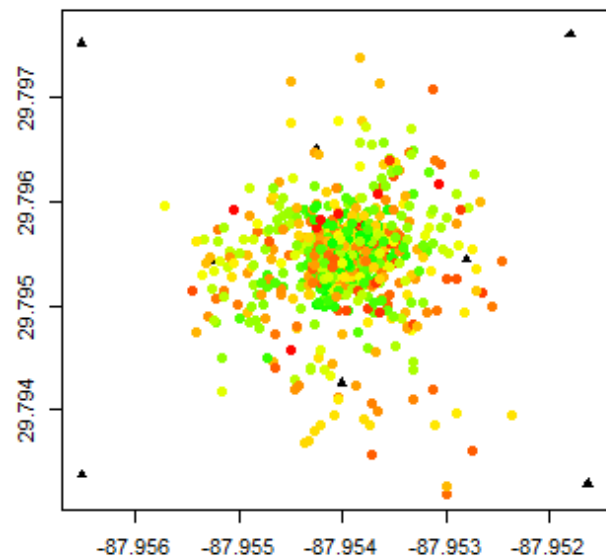
	Pyramid 26	Pyramid 28
Tagged (dart)	5	3
Tagged (acoustic + dart)	18	20
Detected	12	19
Positions	12	17
Stationary and/or outside array	4	6
Moving (low persistence)	1	0
Moving (moderate persistence)	2	4
Moving (high persistence)	5	7

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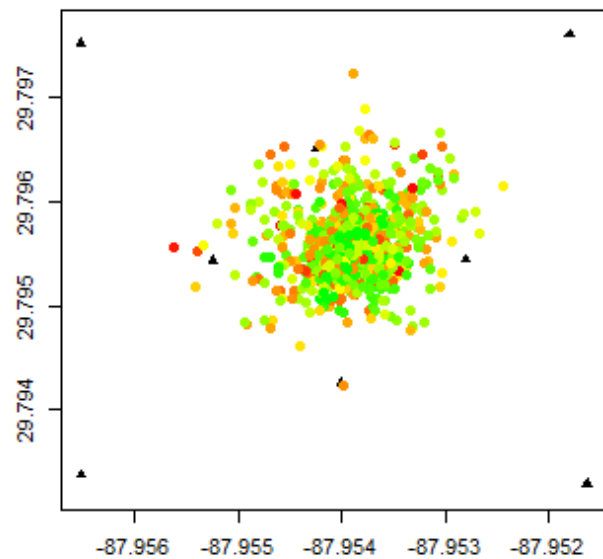


Latitude

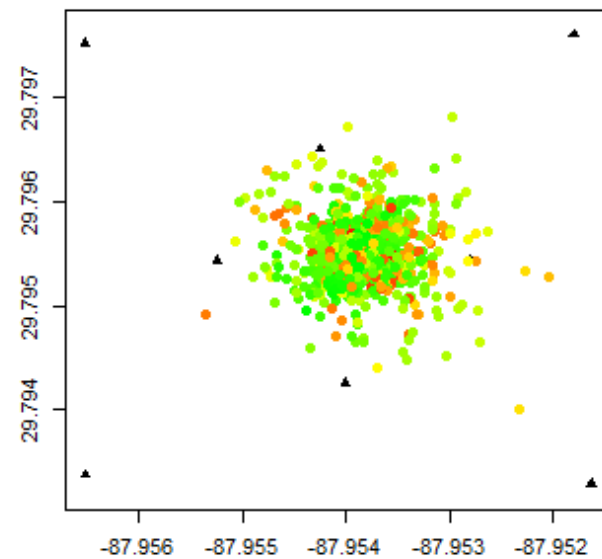
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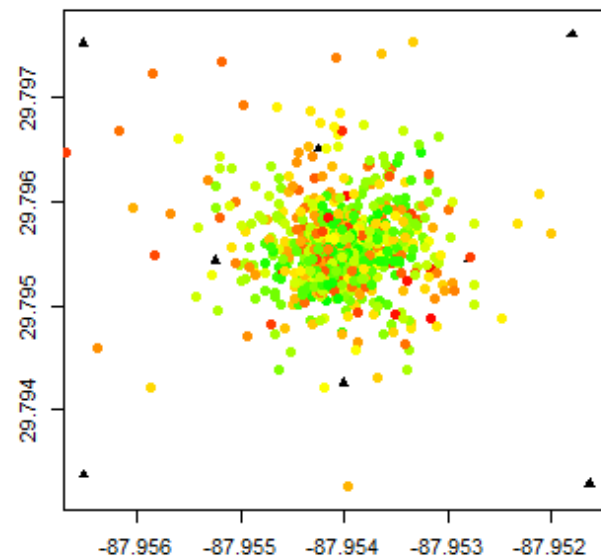
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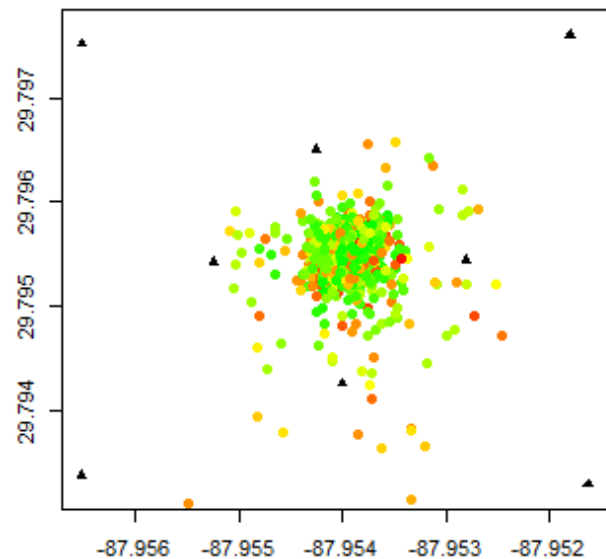
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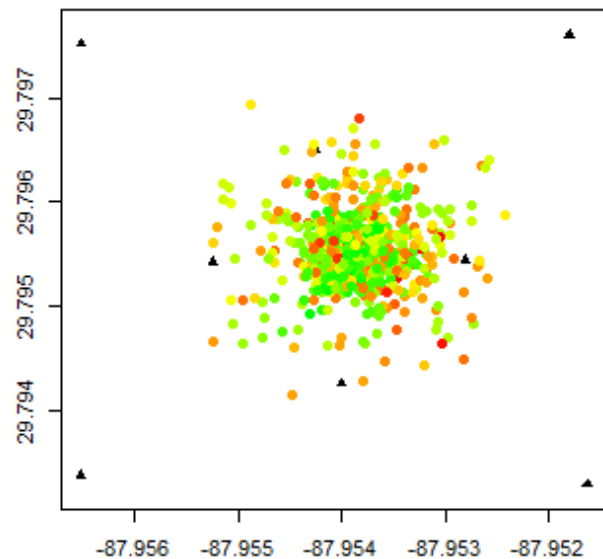
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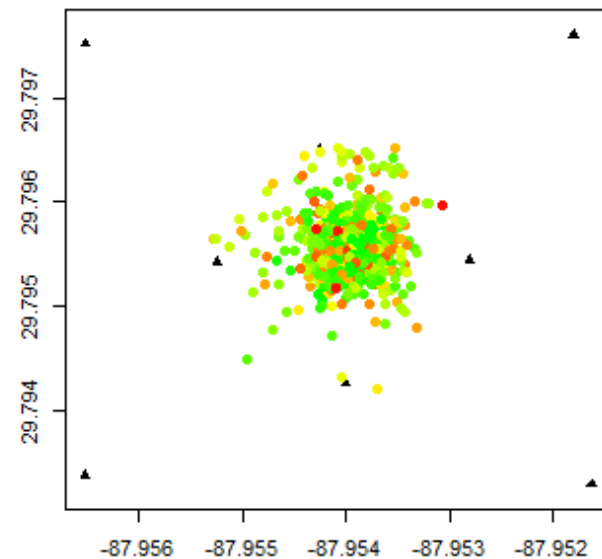
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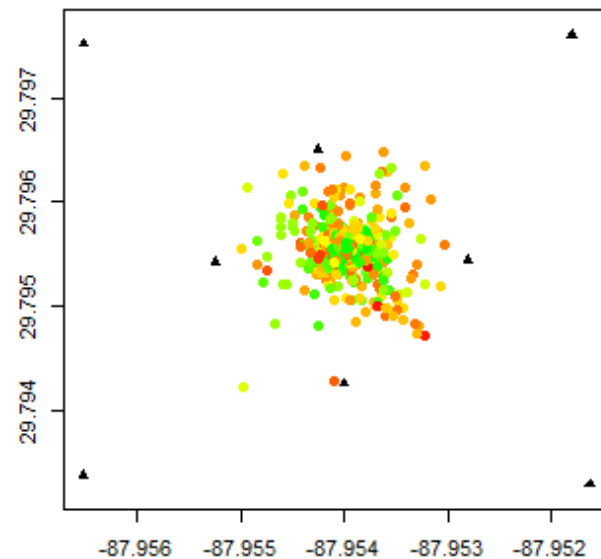
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Longitude

# Lincoln-Petersen density estimate

- Standard L-P mark-recapture density estimator

$$N = \frac{nK}{k}$$

...where  $n$  is the number of fish tagged,  $K$  is the number of fish recaptured and  $k$  is the number of recaps that were tagged

- Assumes that system is closed, so no tagged fish die or leave system between tagging event and recapture event

# VPS L-P density estimate (Shertzer et al, 2020)

- Use acoustically tagged fish to estimate loss factor (combined effect of emigration, mortality, etc.) for all tagged fish
- Apply this factor to number of fish initially tagged to get estimate of number of tagged fish at time of recapture event

$$n = n_a + n_d$$

...where  $n_a$  is the initial number of acoustically tagged fish and  $n_d$  is the initial number of dart tagged fish

$$n' = n'_a + \frac{n_d n'_a}{n_a}$$

...where  $n'_a$  is the number of acoustically tagged fish present based on the VPS position data and  $n'$  is the new estimate of the total number of tagged fish present during a recapture event

- Then use this estimate of tagged fish present during the recapture event in the L-P density estimator...

$$N' = \frac{n'K}{k}$$

...to estimate  $N'$  or the number of fish present during the recapture event

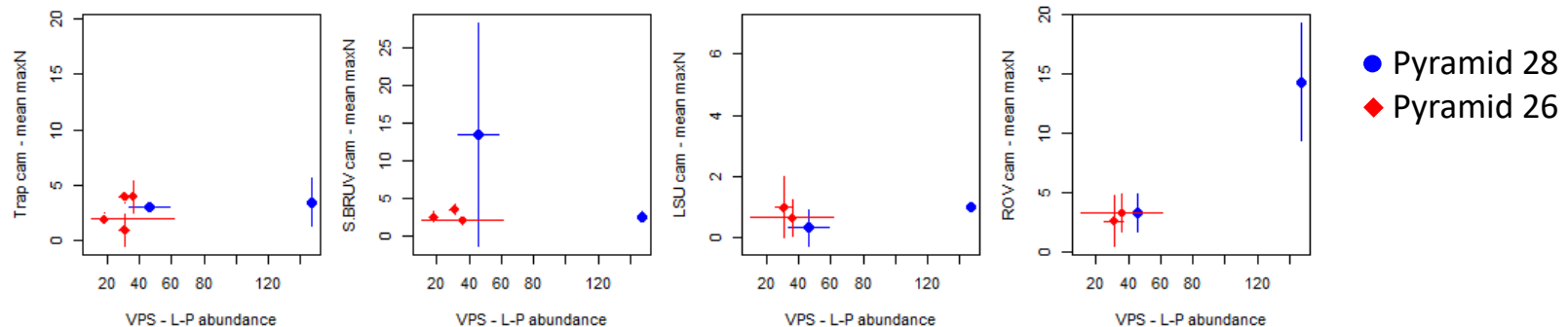


# VPS L-P density estimate

- Few samples where tagged fish were observed; highest number of tagged fish was 1
  - ROV: 5 of 13
  - LSU cam: 0 of 14
  - Trap cam: 2 of 29
  - S-BRUV: 2 of 31

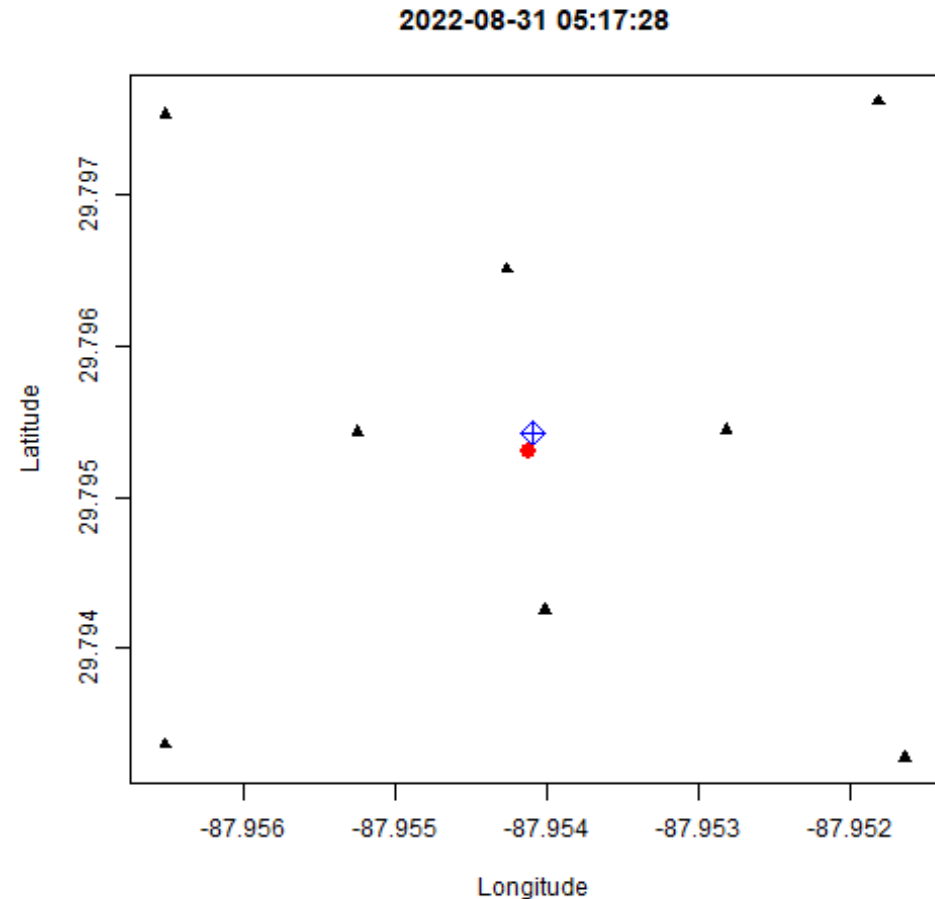
# VPS L-P density estimate

VPS L-P estimates		
	Pyramid 26	Pyramid 28
8/29	31 (S-BRUV)	
8/30	54 (ROV); 18 (S-BRUV)	
8/31		37, 55 (ROV)
9/01	35 (ROV); 27 (Trap)	
9/02	18 (Trap)	147 (ROV)



# VPS behavioral response to gears

- Analyze changes in behavior during gear deployments
- Changes in step length and direction before, during, and after deployment of different gears
- Estimate gear-induced change in density
  - More relevant for continuous vs. discrete habitat patches



# Active acoustics: calibration

## Objectives:

- Test abundance estimation
- Characterize wideband response
- Optimize survey design

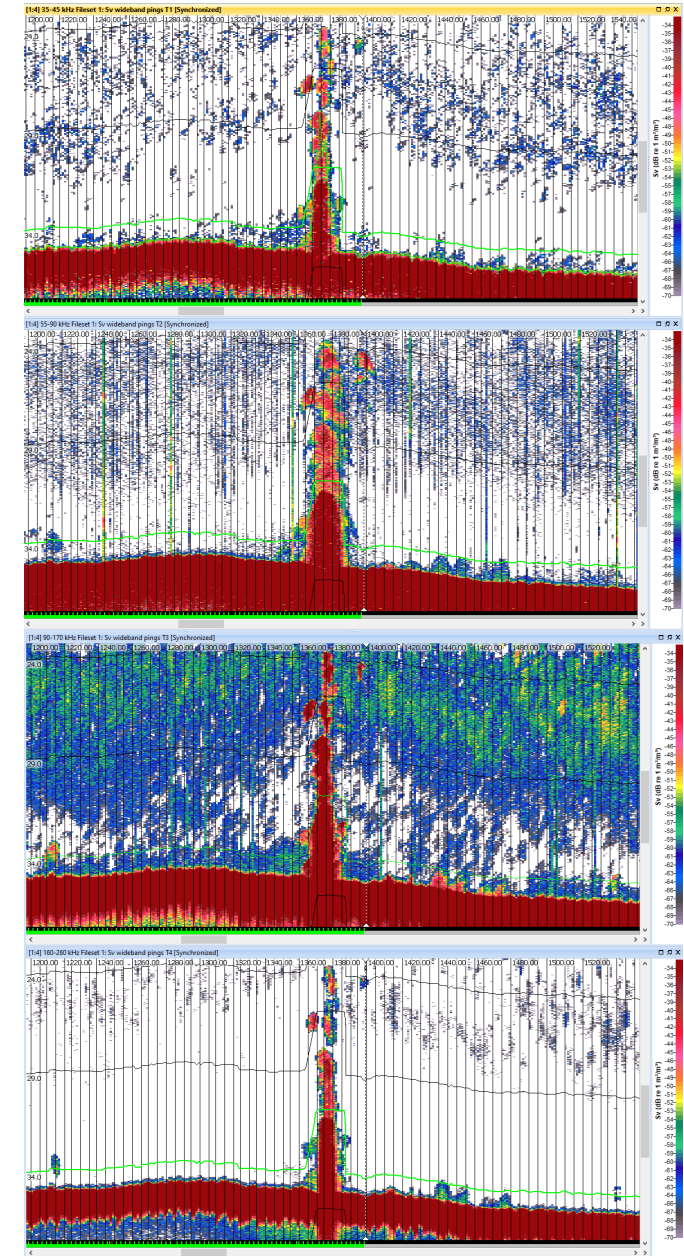
## Data collection:

- Completed one of each survey type each day (3 on SP28, 2 on SP26)
- Four frequencies treated independently
  - 38 (35-45 kHz)
  - 70 (45-90kHz)
  - 120 (90-170kHz)
  - 200 (160-260kHz)



# Active acoustics

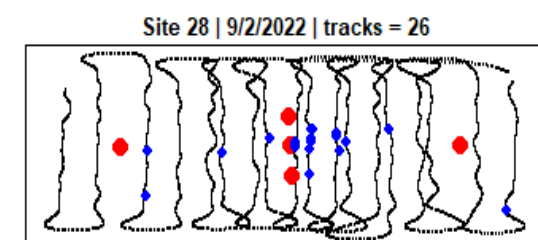
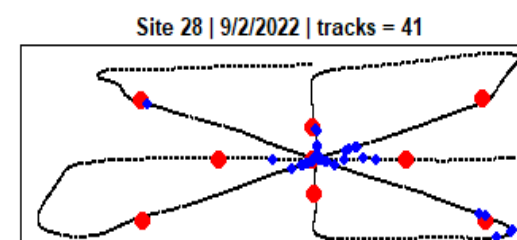
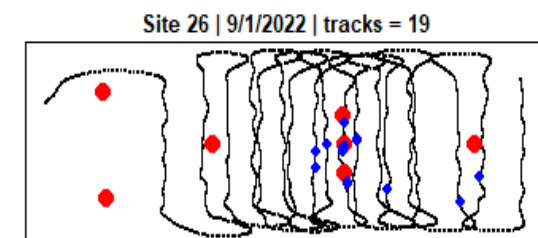
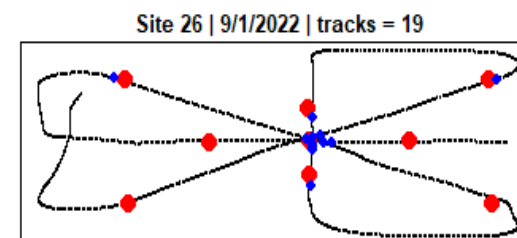
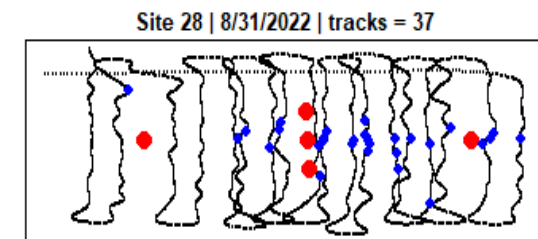
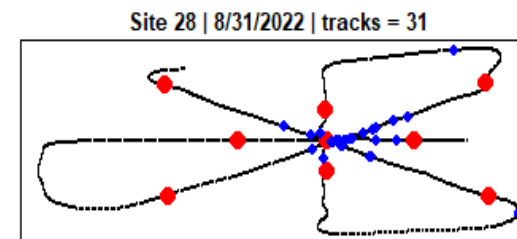
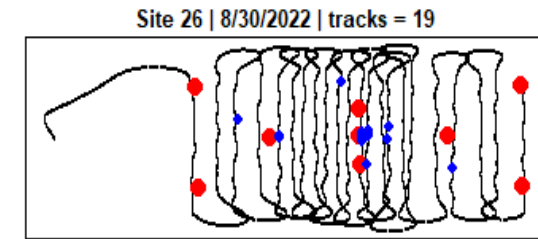
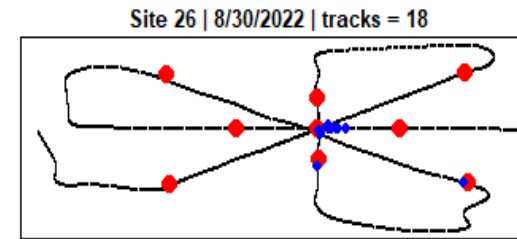
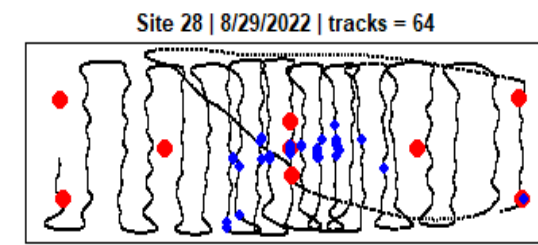
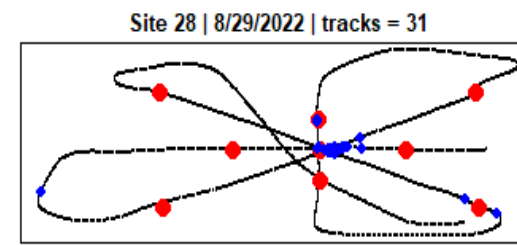
- Beam angle
  - Interaction with depth to determine beam width
  - Can also affect interference related to structures
- Frequency
  - Depending on acoustic signatures, determines ability to observe targets
  - Higher frequencies have higher bandwidth
    - Detect wider range of target types
    - Cost: reduced operational depth
- Results of CT scans combined with calibration results will help us optimize





# Active acoustics

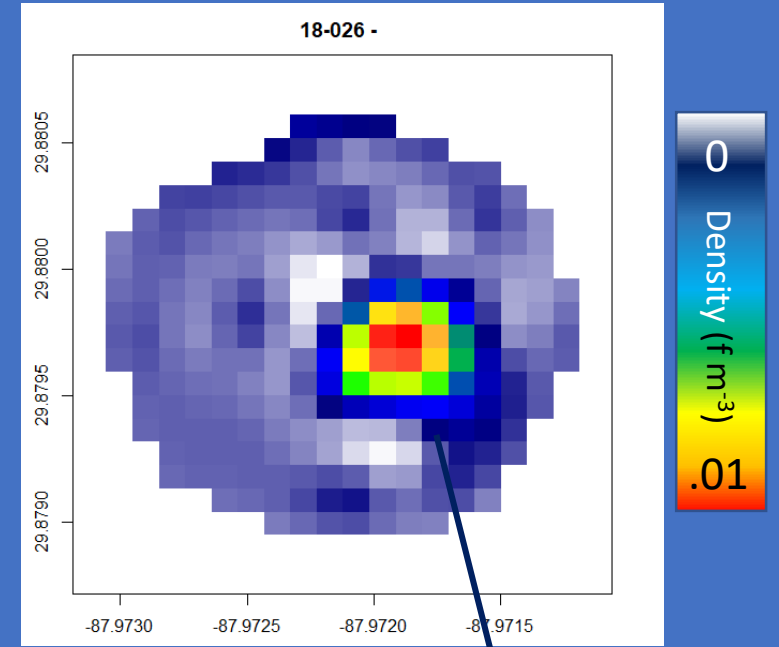
- Fish track counts variable but within reasonable range (18-64 fish per survey)
- Need spatial model to interpolate for density estimates



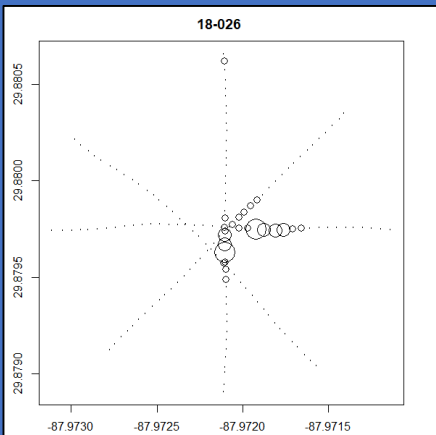
# Spatial Modeling for Abundance Estimation:

- Considered kriging – exponential decay – GAM
- GAM shown to perform well on isolated structures and continuous reefs
- Evaluated across all 4 transducers independently
- Estimates of density ( $\text{f m}^{-3}$ ) are scaled to survey volume for abundance

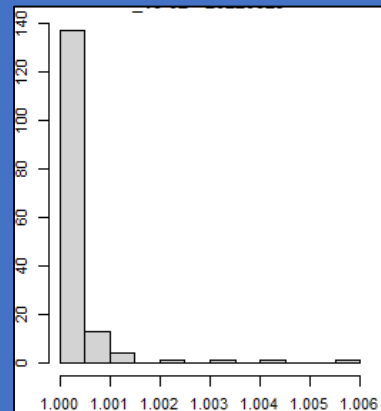
Density predicted for each cell in grid



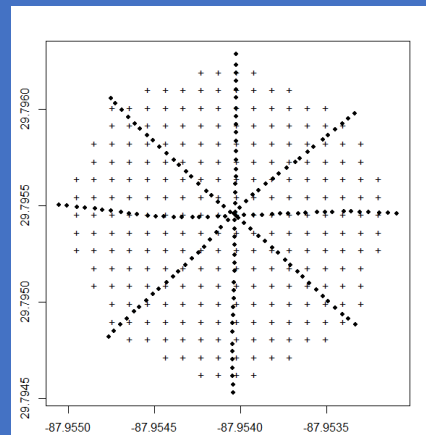
Density evaluated  
in 5x5m cells  
(#fish / cell volume)



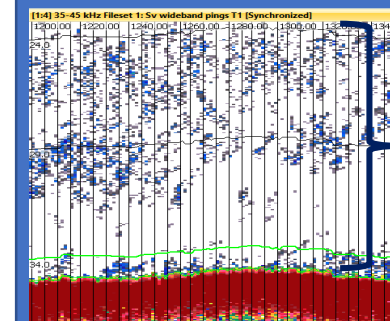
GAM used to  
model fish density:  
 $y \sim s(\text{Lat}, \text{Lon})$   
Tweedie and Gamma\*



Gridded convex hull  
generated for  
predictions



Depth x cell area x  
density = cell  
abundance

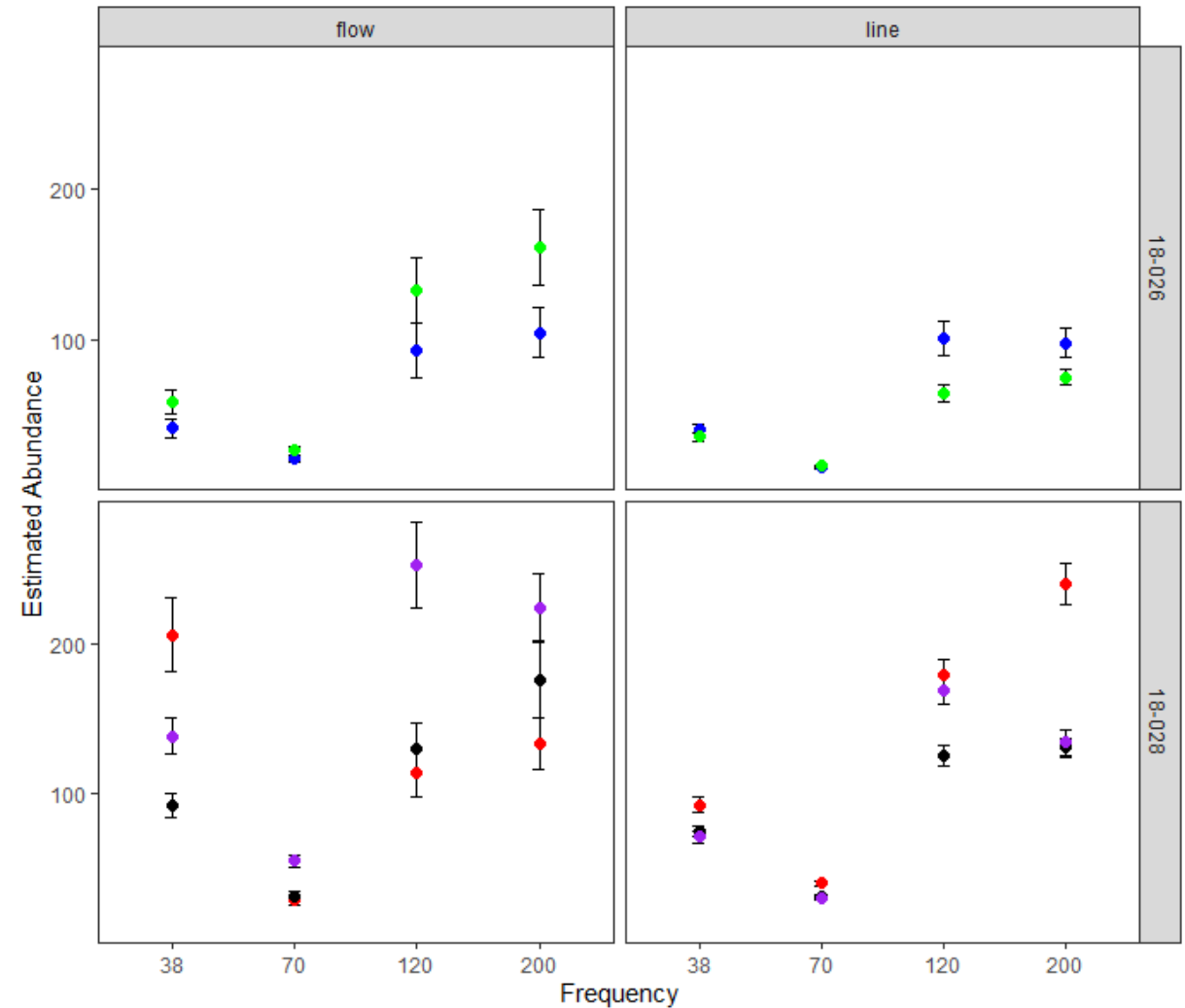


Abundance Estimate  
= sum of cell  
abundance

\*CV estimated from  
model predictions

# Active acoustics

- High variability in predicted density among frequencies
  - Interplay between detectability and beam angle (volume sampled)
- Weak correlations between predicted density and ROV counts, except for 120 kHz
- Preliminary results for 70 kHz echosounder are similar to those from the VPS L-P abundance estimates
- Parallel lines give similar results to flower survey, but with substantially lower variance (but note that the total area covered was higher for parallel lines)



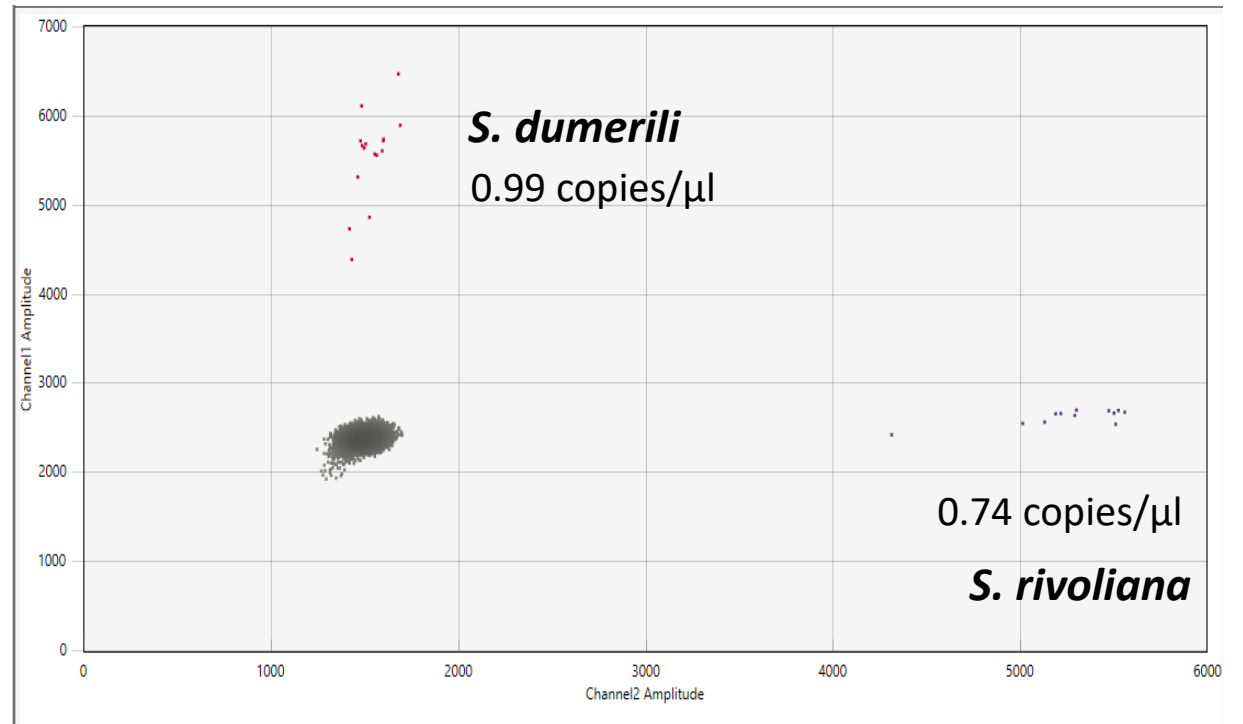
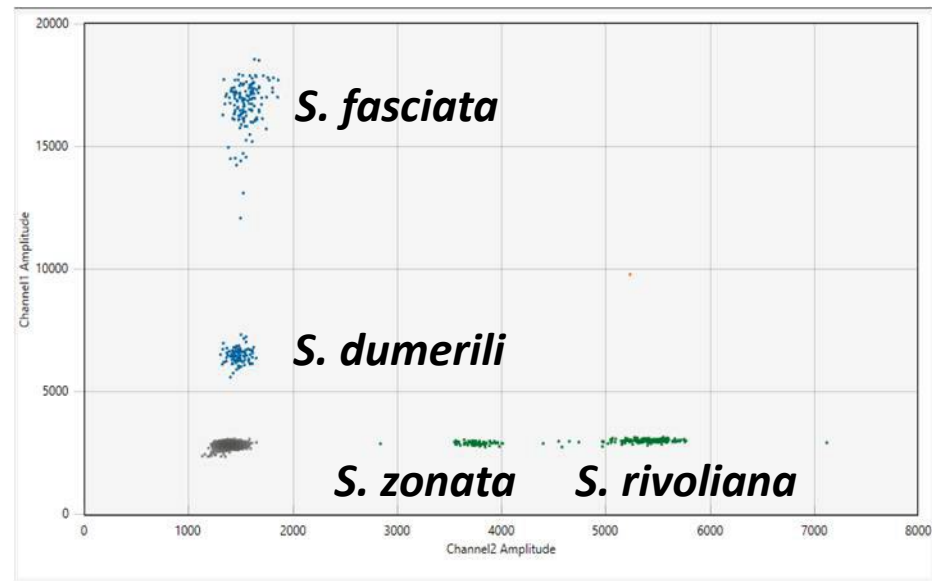
# Active acoustics

## Next steps:

- Standardize beam angle across frequencies to isolate “beam volume dependent detectability” observed in analysis
- Evaluating alternative spatial models
- Calibrate against camera gears across a wider range of fish densities

# eDNA

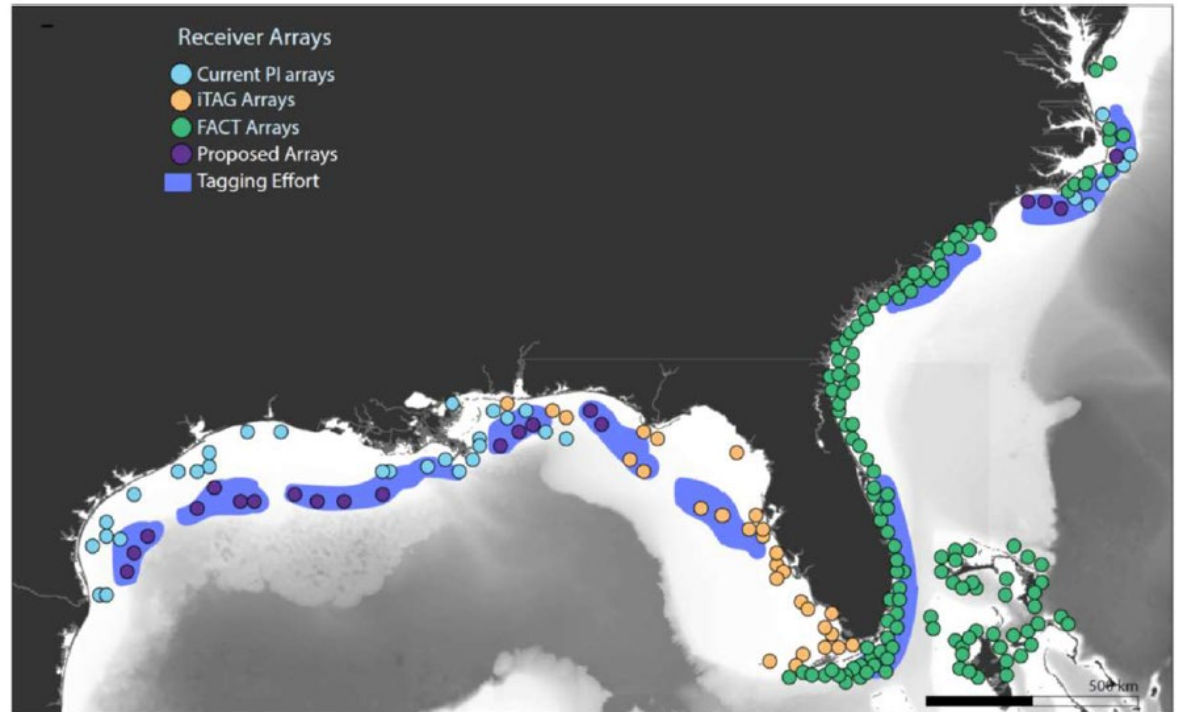
- Of six processed samples (from one day at each site)
  - 4 positive for *S. dumerili*
  - 3 positive for *S. rivoliana*
- Plans to increase detectability
  - Reduce filter pore size
  - Sample downcurrent of site
  - Increase replicate samples
  - Improve cost efficiency





# Objective 4: Movement, connectivity, & mortality

- GoM and SA managed as separate, non-mixing stocks, but little known about migratory behavior and population connectivity
- Combined strategy:
  - Internal acoustic tags + extensive receiver array
  - High-reward external tags
  - Population genetics
- Opportunity for angler engagement



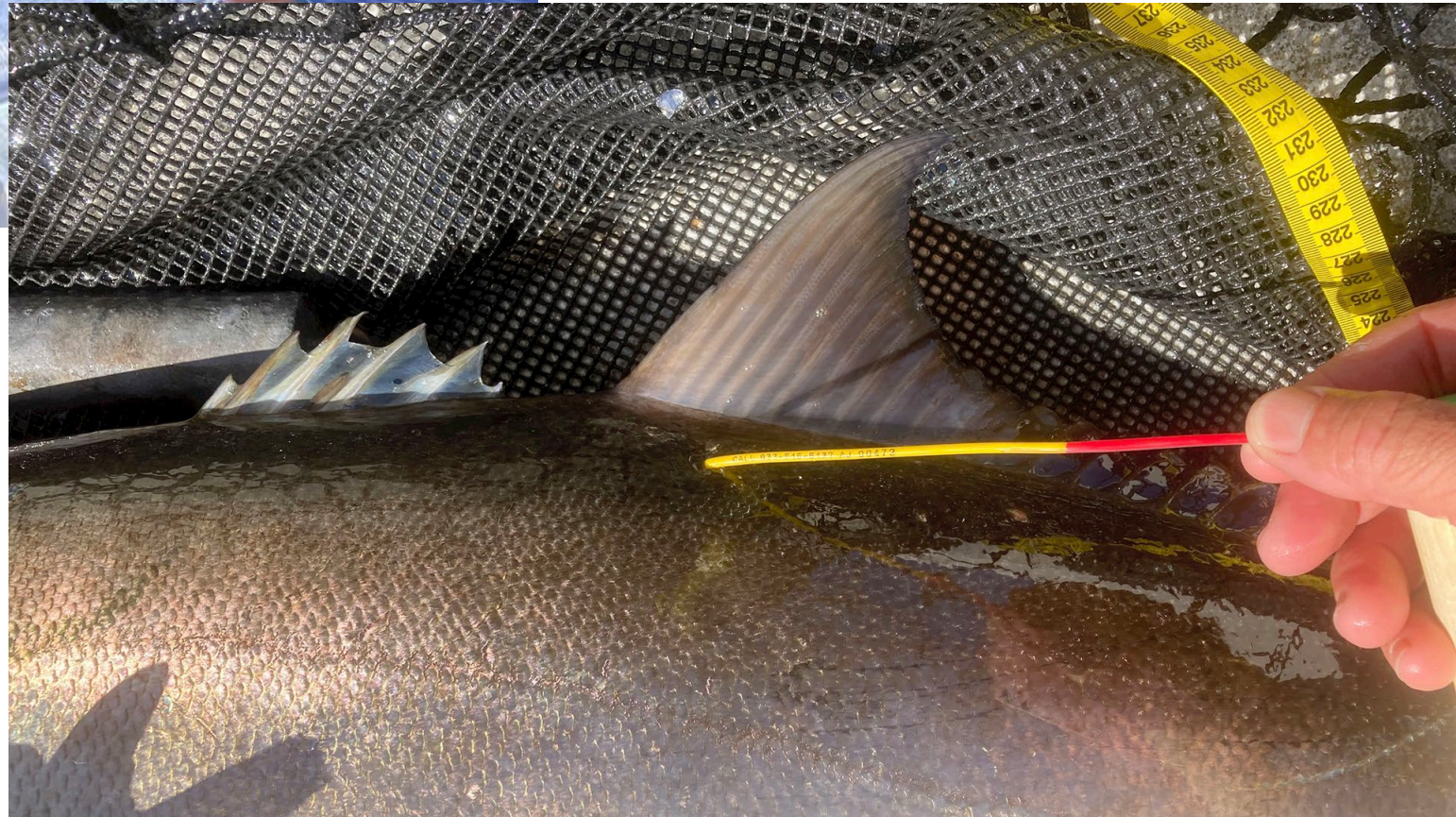




**\$250**

**1-833-515-5137**

[https://agriculture.auburn.edu/research/  
faas/quantitative-fisheries-lab/greater-  
amberjack-project-tag-reporting/](https://agriculture.auburn.edu/research/faas/quantitative-fisheries-lab/greater-amberjack-project-tag-reporting/)





# Conventional tagging

## Objectives:

- Estimate the regional and sector specific (commercial, recreational) fishing mortality rates of Greater Amberjack in the Atlantic Ocean and the Gulf of Mexico
- Assess length-based vulnerability to capture, harvest, and discard
- Evaluate rates of movements of Greater Amberjack among regions

**\$250 Reward for Greater Amberjack Tags**  
Where: Gulf of Mexico and South Atlantic  
Call 1-833-515-5137 to report your tag!

Some of the information we will be looking for:

• Tag number	• Location	• Recreational or Commercial
• Length/Weight	• Date	

**You must clip off all colored external tags and mail them in to receive your \$250 reward\*\***

External Tag:  
Clip off the tag close to the fish

Sea Grant

Tag Return Center Hotline:  
1-833-515-5137


Internal Acoustic Transmitter\*  
Do not disturb if releasing the fish.  
Please save if discovered while filleting.

Sublegal fish have also been tagged. If you catch a sublegal tagged fish, or any tagged fish out of season, please cut the external tag off, release the fish, and call it in.

Tagging data will allow us to estimate movements and fishing mortality rates of Greater Amberjack. Angler tag reporting is essential for this research!

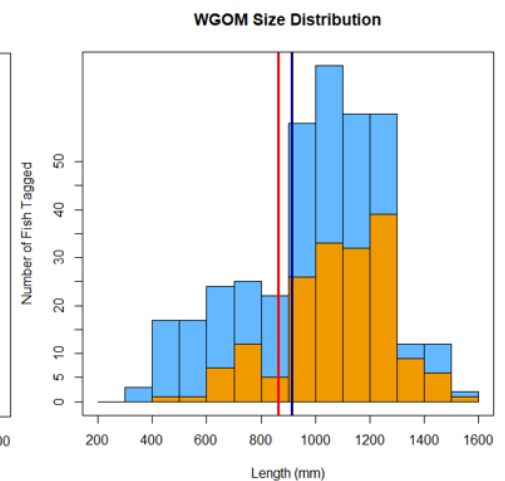
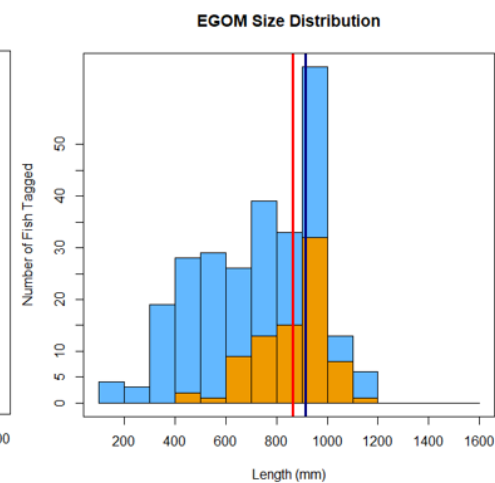
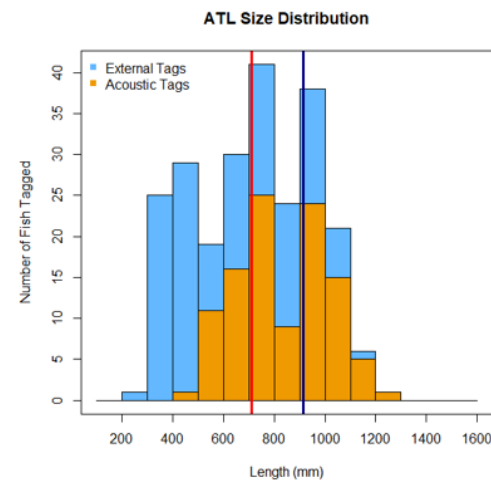
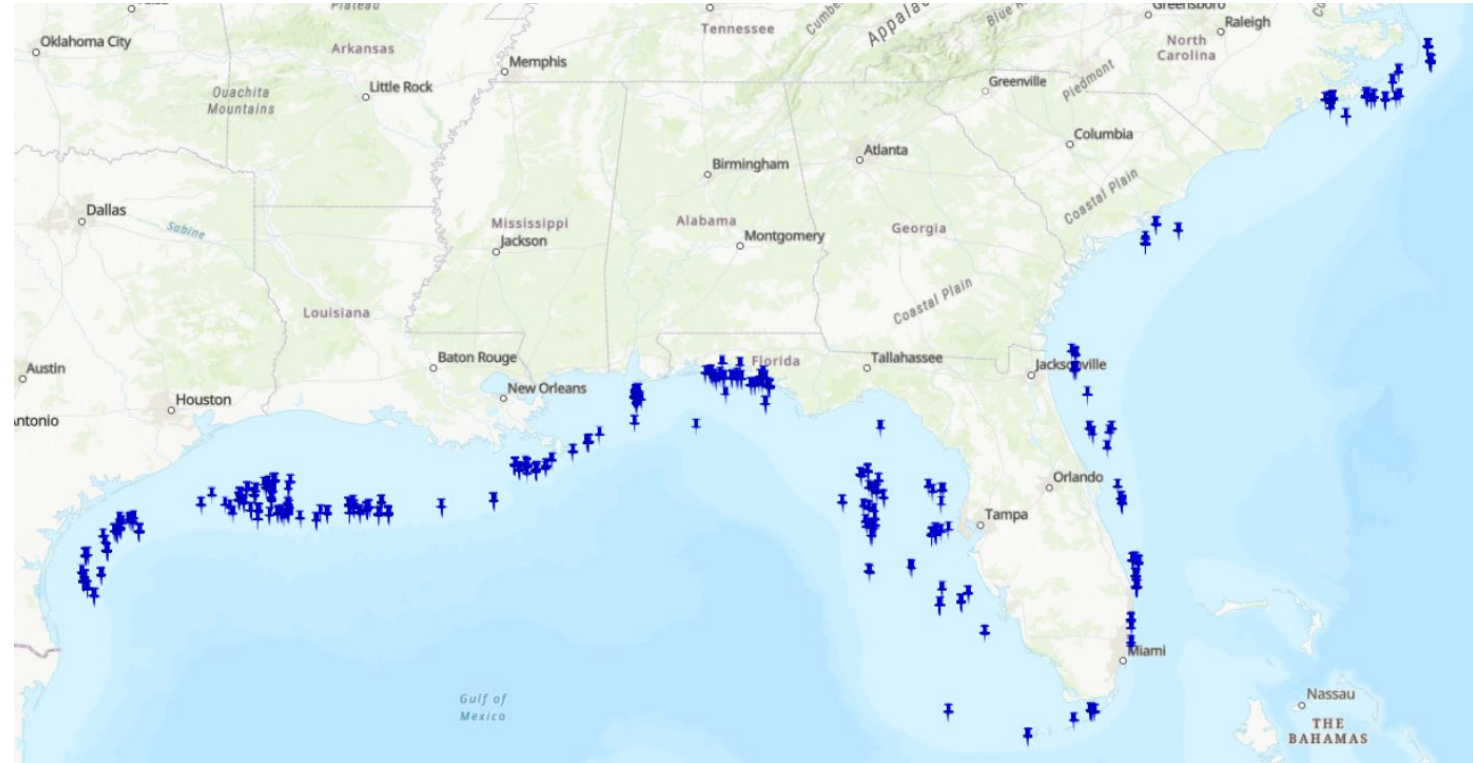
\*Return of transmitter NOT necessary to collect reward.  
\*\*Double-tagged fish receive a single \$250 reward.  
Please clip and send both external reward tags.

AUBURN



# Conventional tagging

- 948/1175 conv. tags out
- 381/336 acoustic tags out
- \$250 reward
- Total tag returns: 72
  - ATL: 21
  - EGOM: 30
  - WGOM: 21
- 7/33 shed tags
- Remaining tags out before beginning of season (Aug 1, 2023)
- Build Bayesian multi-state mark-recapture model
- Incorporate acoustic tag data





# Acoustic tagging

## Objectives:

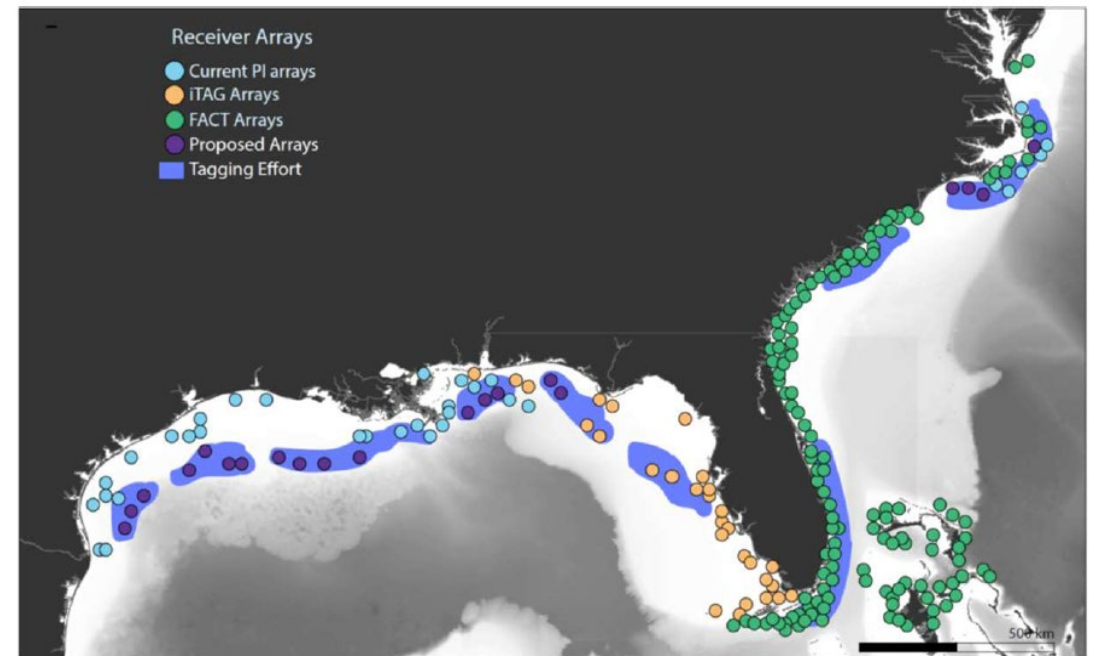
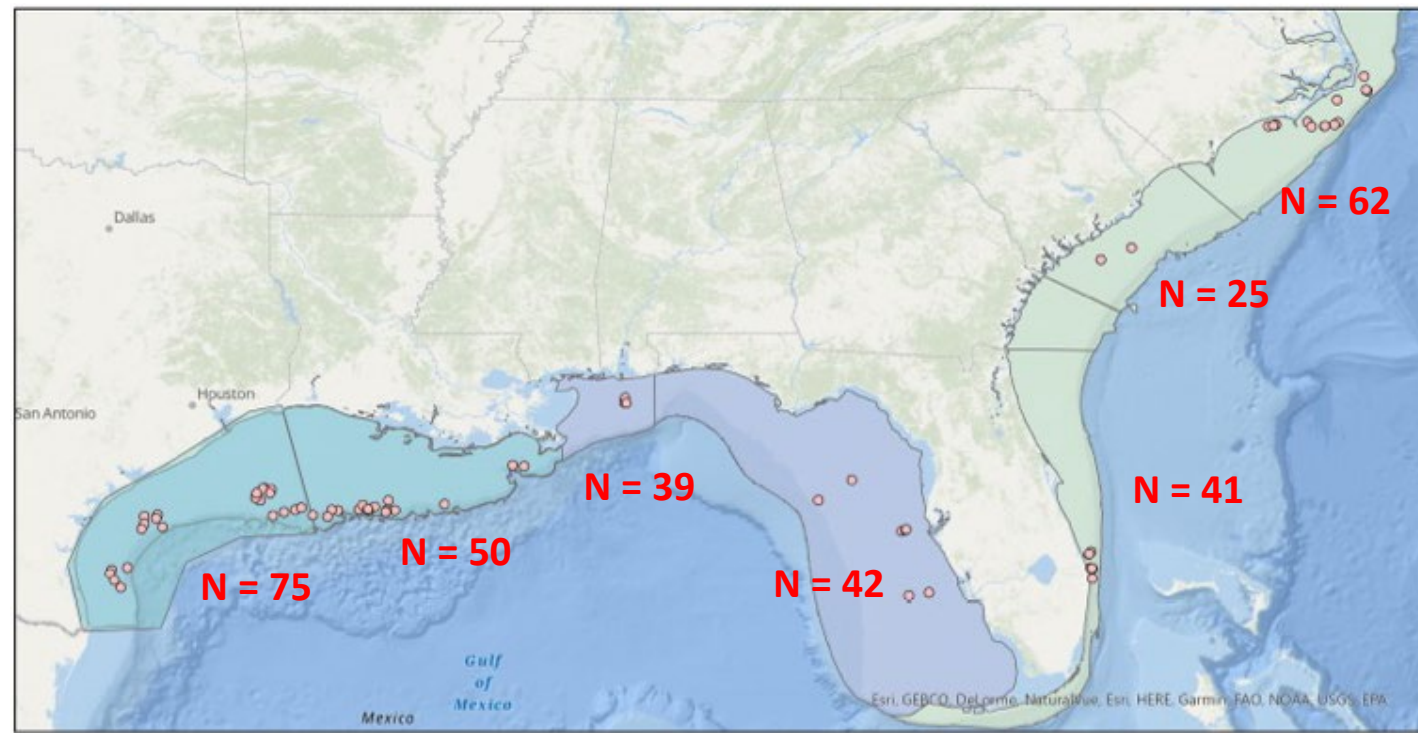
- Residency period/site fidelity by region, structure type, fish size
- Estimates of movement and exchange within and between regions (SA, EG, WG)
- Estimate mortality (F and M)
- Post-release mortality estimates
- Depth use across habitat types and regions





# Acoustic tagging

- 381/336 tags out
- Coordination with iTAG and FACT
- Receiver downloads Summer/Fall 2023



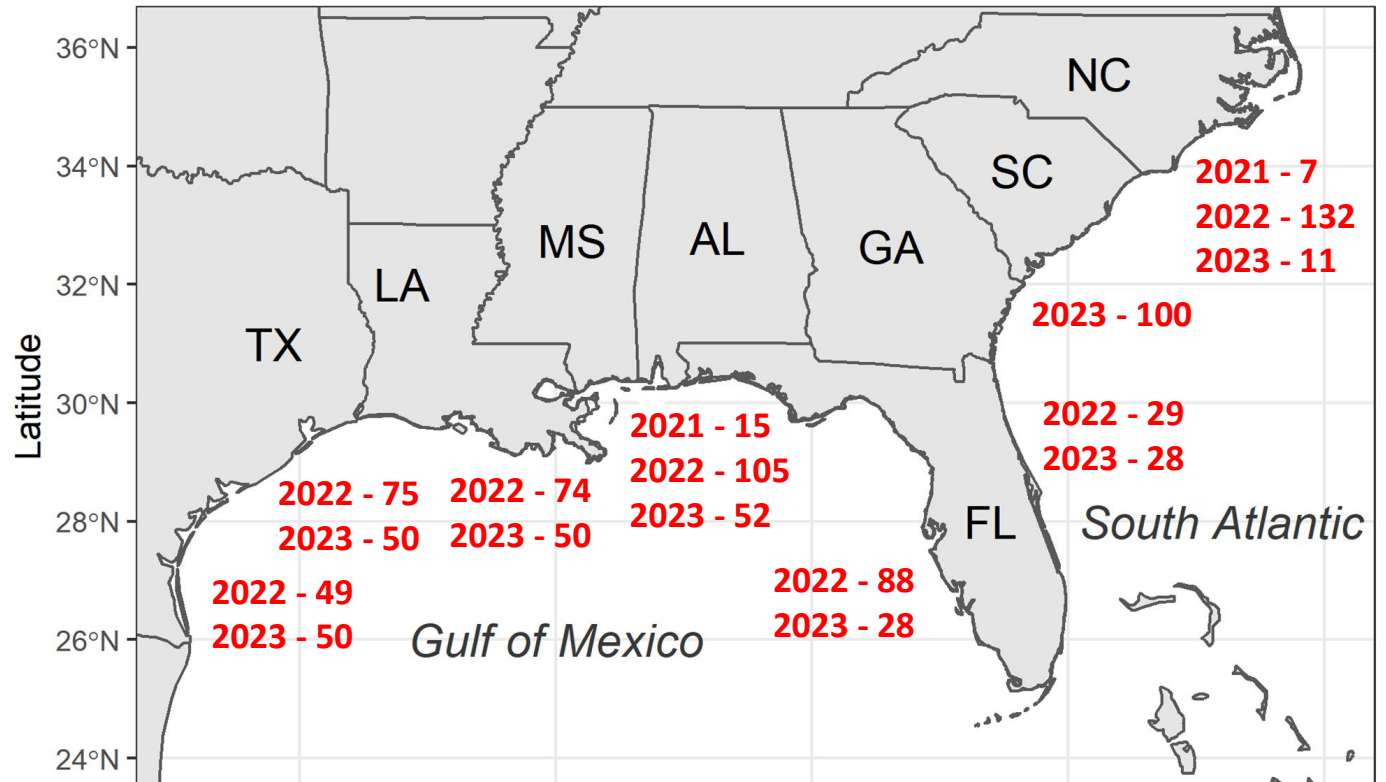
# Population genetics

## Objectives:

- Develop genomic resources to interpret genome scans in greater amberjack
  - Draft genome assembly
  - Linkage map
- Survey population genetic structure in GoM and SA waters
  - Sample geographic populations and assay samples at 2,000 to 10,000 SNP
  - Analyze genetic stock structure and connectivity: identify units, infer migrants and migration patterns, analyze variation under selection

# Population genetics

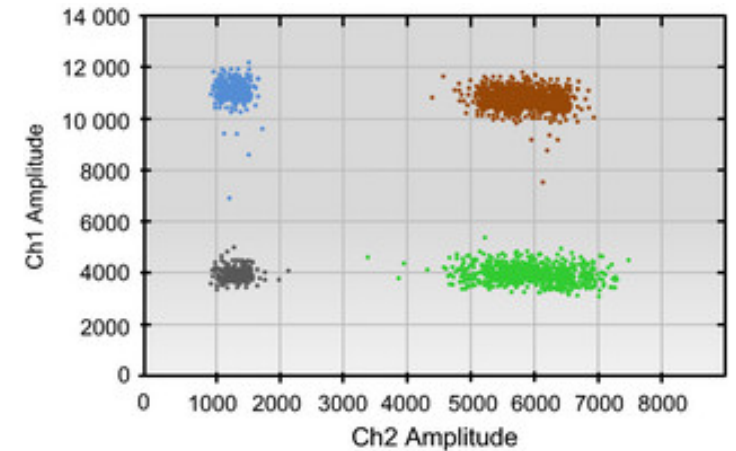
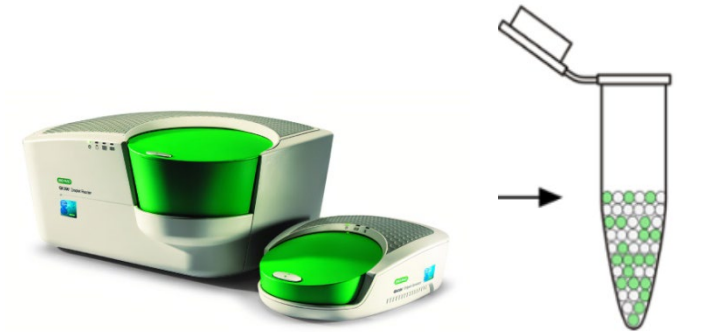
- Progress on reference genome
- Sample population
  - Tagging project
  - Fishery dependent
- To do:
  - Complete reference genome
  - Complete linkage map
  - Assay population sample using dd-RAD sequencing
  - Analyze genetic stock structure and connectivity
- Note: samples archived for future analysis (parentage)



**We also have access to samples from several non-US locations that will be used as reference**

# Objective 5: Environmental DNA

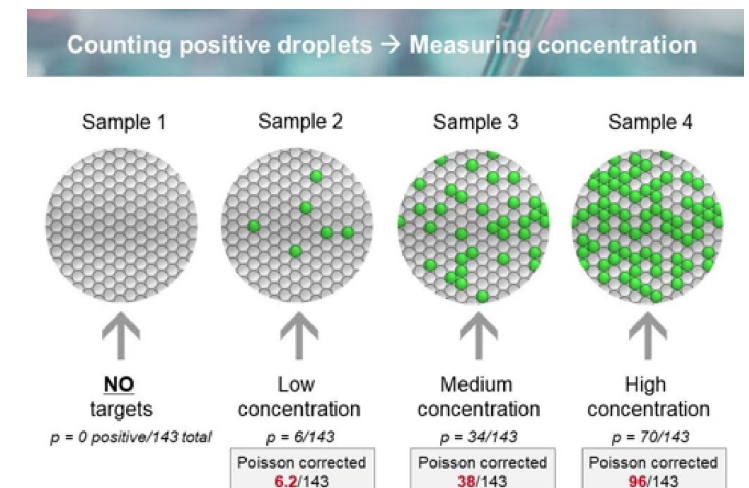
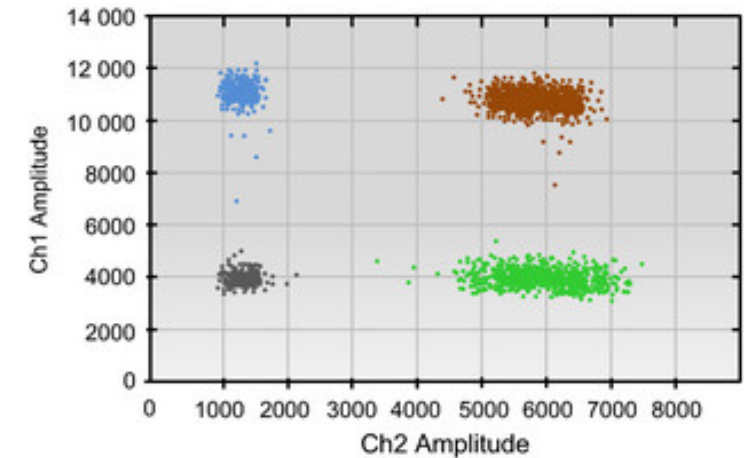
- Investigate efficacy of, and use, eDNA to assess presence and relative abundance of GAJ and closely related species
- Develop novel eDNA tools (ddPCR assay) specific to GAJ and compare performance to other gears during calibrations and regular surveys
  - Confirm identification of species
  - Estimate “sampling” vs. “structural” zeros
  - Provide relative abundance estimates
- Proving ground for the use of eDNA tools to study distribution and abundance of marine fishes



# eDNA

## Objectives:

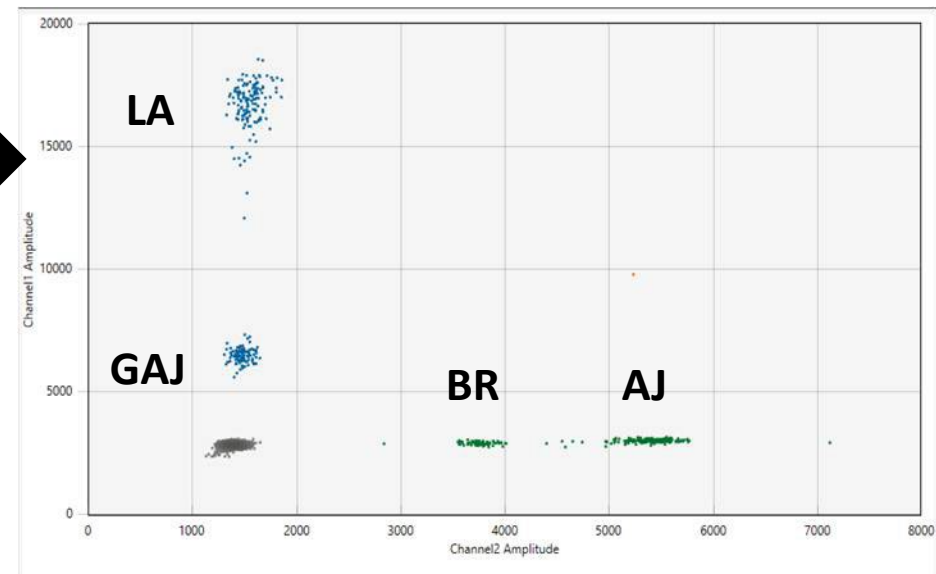
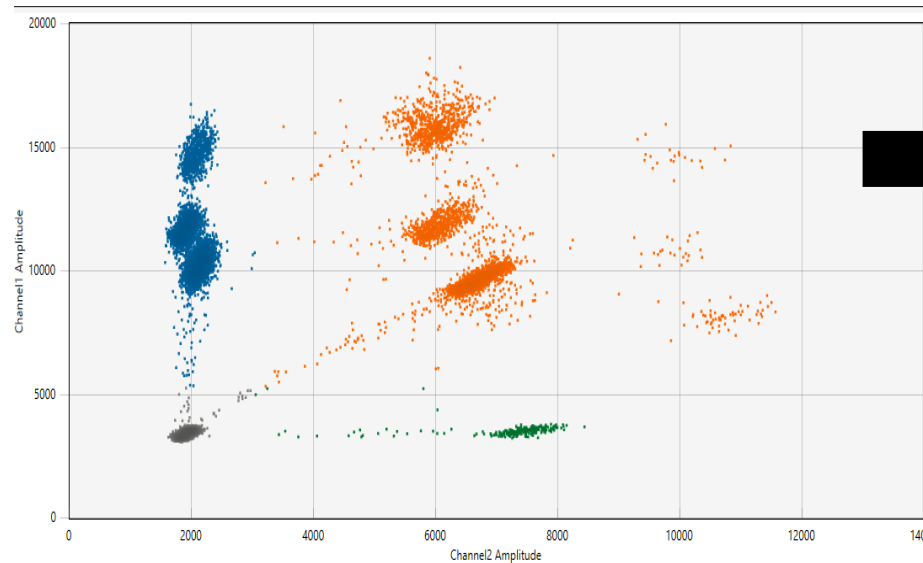
- Evaluate capacity for eDNA tools to detect, discriminate and quantify target DNA
- Develop ddPCR assay
- Work out sampling tools and techniques for system
- Collect field data in concert with other gears
  - Calibration
  - Abundance sampling





# eDNA assay

- Four probes
- > 10 combinations tested
  - ddPCR conditions optimized
  - Cross-test on 24 non-target species including bait and other *Seriola* spp.



# eDNA sampling

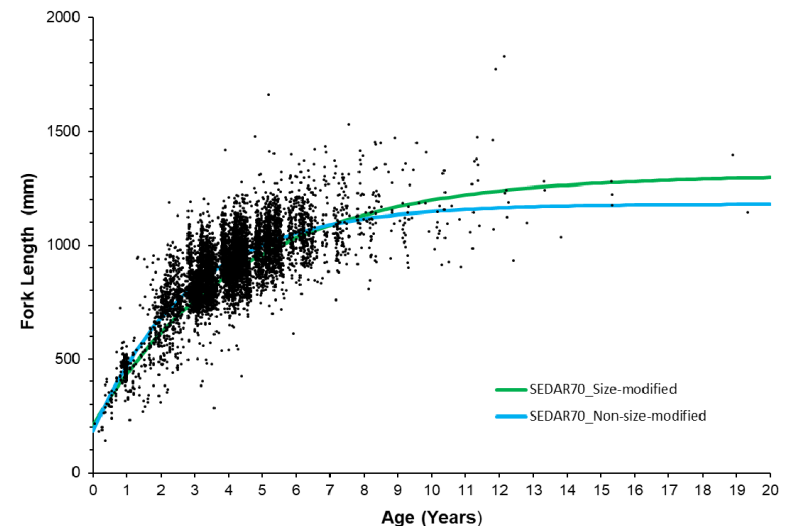
- Methods
  - Triplicate parallel Niskin samples
  - Triplicate serial Niskin samples
  - Passive samplers (sponges) mounted on ROV
- Preliminary results
  - Niskin samples appear to miss fish when present
  - Passive samplers mounted to ROV more effective at detecting fish when present
  - Need to run more samples from places where fish not observed on camera gears



Passive samplers

# Objective 6: Update biological information

- Recent stock assessments recommended expanded demographic sampling of GAJ
- Age and growth information from W-GoM has been extremely limited
- Will use fishery dependent and fishery independent collections to update biological information and refine age-length keys
- Archive samples that can be used (with additional funding) to update reproductive indices (fecundity, spawning season, etc.)



# Objective 7: Stakeholder engagement

- Working closely with established groups (e.g., GAJ Visioning Team, Sea Grant Reef Fish Extension Collaborative, etc.) to facilitate communication and cooperation with stakeholders
- Start-to-finish:
  - GAJ Visioning Team collected stakeholder input – used to formulate goals of RFP – Funded research is responsive to priorities of RFP
  - Incorporation of LEK in study design
  - Active engagement with for-hire fishing sector to provide platforms for scientific sampling
  - Dependent on commercial and recreational anglers for high-reward conventional tag returns
  - Dedicated effort to communicate results broadly at conclusion of study

# Expected impacts and application of results

- Large-scale survey using novel integrated sampling approaches
- Leverage existing data sets and ongoing research to augment data collection and cost effectiveness
- Primary benefits:
  - Robust estimate of absolute abundance of age 1+ GAJ in GoM and SA
  - Improved understanding of spatial and habitat-related distribution of the species
  - Improved understanding of population and movement dynamics of GAJ in region
  - Development of an approach and analysis framework that can be applied to future GAJ abundance estimates and those for other reef-fish species
- Secondary benefits:
  - Estimates of GAJ growth, mortality, site fidelity, population connectivity
  - Improved understanding of reef fish community structure across study region

A large school of yellow-striped snappers (Lutjanus fulvus) is swimming in clear, deep blue water. The fish are silvery with a distinct yellow stripe running along the side of their bodies. They are arranged in a loose, coordinated pattern, moving towards the right side of the frame. The lighting is bright and even, highlighting the sleek, elongated shapes of the fish and their pointed snouts. The background is a uniform, vibrant blue, suggesting a healthy, open-ocean environment.

Questions